

LYONS FERRY COMPLEX ANNUAL OPERATION PLAN

For the Period of

OCTOBER 1, 2007 – SEPTEMBER 30, 2008

Prepared by:

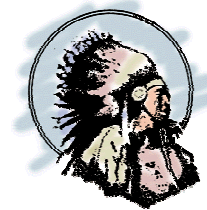
Washington Department of Fish and Wildlife



Nez Perce Tribe



Confederated Tribes of the
Umatilla Indian Reservation



And funded by:

Lower Snake River Compensation Plan



TABLE OF CONTENTS

I. INTRODUCTION	1
A. Facilities	1
1. Lyons Ferry Hatchery	2
2. Tucannon Hatchery	2
3. Cottonwood Acclimation Facility	3
4. Dayton Acclimation Facility	3
5. Curl Lake Acclimation Pond	4
6. Other Acclimation Facilities	4
B. Fish Production Summary	4
II. SNAKE RIVER FALL CHINOOK	7
A. Fish on Hand	10
B. Trapping	10
1. Lyons Ferry Hatchery	10
2. Lower Granite Dam	10
C. Spawning	11
D. Rearing	12
E. Tagging, Transfers, and Releases Assuming Full Production	13
F. Research	15
III. TUCANNON SPRING CHINOOK	16
A. Fish on Hand	16
B. Trapping	16
C. Spawning	17
D. Rearing	17
E. Tagging, Transfers, and Releases	17
F. Research	18
IV. SUMMER STEELHEAD - GENERAL	18
V. LYONS FERRY SUMMER STEELHEAD	19
A. Fish On Hand	19
B. Trapping	19
C. Spawning	20
D. Rearing	20
E. Tagging, Transfers, and Releases	20
F. Research	21
VI. TOUCHET SUMMER STEELHEAD	21
A. Fish on Hand	21
B. Trapping	21
C. Spawning	21
D. Rearing	22
E. Tagging, Transfers, and Releases	22
F. Research	22
VII. TUCANNON SUMMER STEELHEAD	22
A. Fish on Hand	23

B. Trapping	23
C. Spawning.....	23
D. Rearing.....	23
E. Tagging, Transfers, and Releases.....	24
F. Research.....	24
VIII. WALLOWA SUMMER STEELHEAD.....	24
A. Fish on Hand	24
B. Trapping	25
C. Spawning.....	25
D. Rearing.....	25
E. Tagging, Transfers, and Releases.....	25
F. Research.....	26
A. Fish on Hand	26
B. Rearing	27
C. Tagging, Transfers, and Releases.....	27
X. FISH HEALTH	28
A. Guiding Policies.....	28
B. Monitoring.....	29
C. Specific Fish Health Management	29
1. BKD Management – Fall Chinook	29
2. IHN Management – Summer Steelhead	29
3. Broodstock and Egg Fungus Management	30
XI. COMMUNICATION.....	31
Appendix A: 2007 Requests for Production Fish/Eggs.....	33
Appendix B: 2007 Fall Chinook Trapping/Sampling Protocol.....	35
Appendix C: 2007 Mating Matrix for Spawning at LFH.....	38
(2007Broodyear).....	38

List of Figures

Figure 1. Map of the Lower Snake River Compensation Plan (LSRCP) LFC Facilities, and major rivers and streams in Southeast Washington.	1
---	---

List of Tables

Table 1. LFC production capacities (historical design versus current).	5
Table 2. LFC plants and transfers by brood years (BY) – three-year profile.	6
Table 3. Revised production table listing Snake River fall Chinook salmon production priorities for LFH as documented in the June 1, 2006 Draft of the SRFMP.	9
Table 4. Identified Areas for fall Chinook juvenile and Adult out planting as presented in the June 1, 2006 Draft SRFMP.	12
Table 5. Proposed 2007 - 2008 Snake River fall Chinook tagging, transfers and releases.	14
Table 6. Proposed 2007 - 2008 Tucannon River spring Chinook tagging, transfers and releases.	18
Table 7. Proposed 2007 - 2008 LFH stock summer steelhead tagging, transfers and releases. .	20
Table 8. Proposed 2007 - 2008 Touchet River summer steelhead tagging, transfers and releases.	22
Table 9. Proposed 2006 - 2007 Tucannon River summer steelhead tagging, transfers and releases.	24
Table 10. Proposed 2007 – 2008 Wallowa stock summer steelhead tagging, transfers and releases.	25
Table 11. 2007 Kamloops rainbow trout tagging, transfers and releases.	28
Table 12. 2008 Spokane rainbow trout tagging, transfers and releases.	28
Table 13. Contact List.	31

I. INTRODUCTION

A. Facilities

Lyons Ferry Complex (LFC; See Figure 1) includes Lyons Ferry Hatchery (LFH), Tucannon Hatchery (TFH), Cottonwood Acclimation Facility (Cottonwood AF), Dayton Acclimation Facility (Dayton AF), and Curl Lake Acclimation Pond (Curl Lake AP).

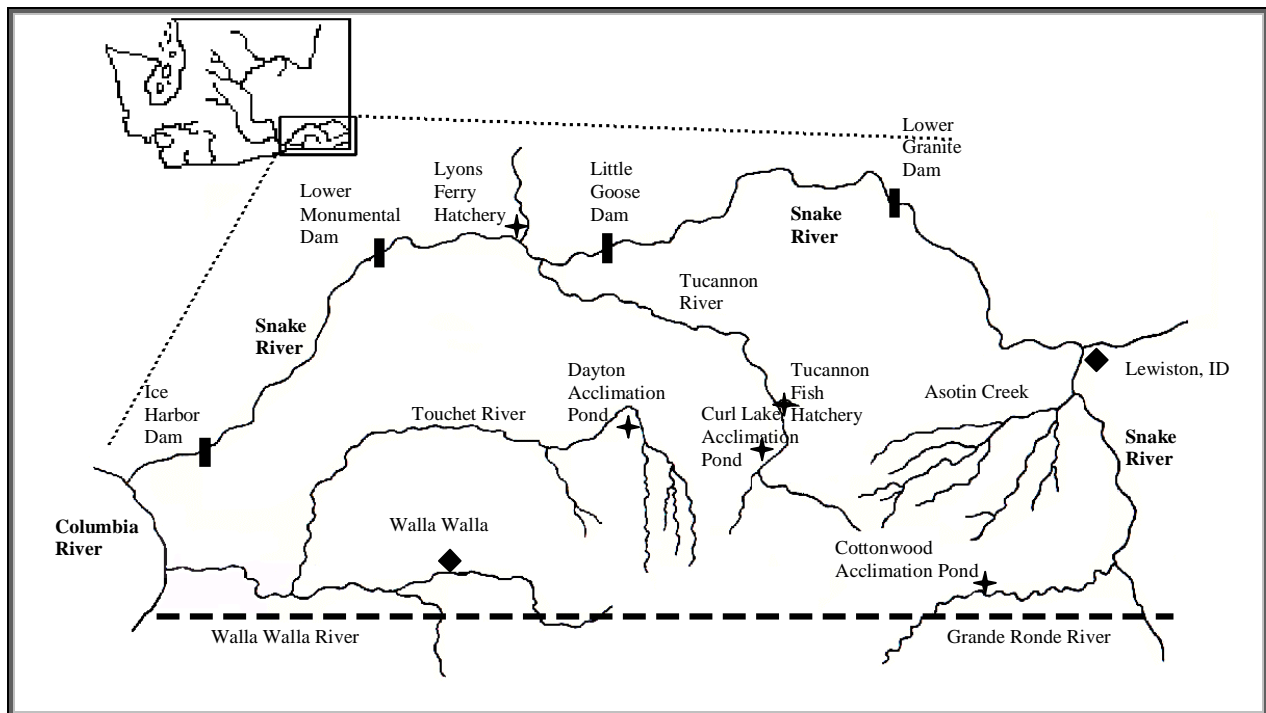


Figure 1. Map of the Lower Snake River Compensation Plan (LSRCP) LFC Facilities, and major rivers and streams in Southeast Washington.

LSRCP funded fish production in Washington began in 1983, with the construction of trout and steelhead rearing facilities at the LFH. Construction of salmon facilities and steelhead acclimation sites followed, and was completed in 1985. Major upgrades at TFH also occurred at that time, and operation of that facility has been funded by LSRCP every since. Production at all facilities has been directed toward meeting established program goals of returning 18,300 adult fall Chinook, 1,152 adult spring Chinook, 4,656 adult summer steelhead, and providing 67,500 angler days of fishing opportunity from 80,000 pounds of rainbow trout production, currently planted at 3 fish per pound (fpp). In addition to these LSRCP production goals, Washington Department of Fish and Wildlife (WDFW) funds a jumbo-sized (1.5 pounds each) rainbow trout program at TFH.

1. Lyons Ferry Hatchery

The LFH is located along the Snake River at river mile (RM) 59.1, directly below the confluence of the Palouse River in Franklin County, Washington. Initially it was operated as two separate facilities. Washington Department of Wildlife (WDW) operated the north hatchery, producing steelhead and rainbow trout. Washington Department of Fisheries (WDF) operated the south hatchery, rearing spring and fall Chinook. A merger of the two agencies in 1994 led to a merging of the two facilities, and has since been operated by WDFW through LSRCP funding as LFH.

Facilities include two incubation buildings with office space and feed storage, plus adult fish trapping, holding and spawning structures. A visitor center provides interpretive information for guests of the hatchery. There are eight residences on-site for staff to fulfill security and emergency response needs.

The LFH rearing facilities include twenty-eight raceways at 10 ft x 100 ft x 2.8 ft and nineteen raceways at 10 ft x 88.5 ft x 3.5 ft. These raceways were covered in 2" square mesh netting in 2005 and 2006. There are three rearing lakes covered in 4" netting (added in 2003-04), holding ~ 590,000 cubic feet (ft³) of water each, approximately 1,100 ft x 90 ft x 10 ft in size. Netting has been added to these lakes and raceways to reduce predation losses. The adult holding facilities include three 83 ft x 10 ft x 5 ft adult raceways with housed spawning facilities incorporated over the center of these ponds, two 18 ft x 150 ft x 4.3 ft and two 21 ft x 150 ft x 4.3 ft adult salmon holding ponds, which also accommodate sub-yearling rearing when not needed for adult holding in the spring of the year. In 2005, channels were cut into two of these ponds, creating three temporary holding areas in each of the two modified ponds to accommodate marking and tagging of the subyearlings reared there. Screens were fabricated to fit the channels. Eight 20 ft x 4 ft fiberglass circular ponds are used for a captive brood spring Chinook program. These ponds were added in 1998 below the north raceways. No longer used for this program are fifteen 4 ft x 1.6 ft fiberglass circular ponds. Six 3.25 ft x 16 ft x 2.6 ft fiberglass tanks were added in the same area in 2006, allowing for decreased densities and improved flexibility in all stocks during early rearing. The incubation facilities include 112 full stacks (2 units of 8 trays each) of vertical incubators in the south trough room, and 88 shallow eyeing/hatching troughs and four 3.75 ft x 27.5 ft x 2 ft intermediate rearing troughs in the north trough room.

Water is supplied to LFH from the Marmes pump station, which has emergency power backup generation. The Marmes pump (wells) facility has three 300 horsepower (hp) pumps, four 200 hp pumps and one 75 hp pump. The well water right for LFH is 53,200 gallons per minute (gpm), or 118.5 cubic feet per second (cfs) of flow, and water temperature is a constant 52° F.

2. Tucannon Hatchery

The TFH is located along the Tucannon River, between the towns of Dayton and Pomeroy Washington, at RM 36 in Columbia County. Fish production began in 1949 by the Washington Department of Game. In 1983, construction began to remodel the hatchery as part of a transfer of ownership to LSRCP. In November 1986 construction was complete, and LSRCP has funded operations there ever since.

The TFH includes a combined incubation and office building, back-up power generation building, feed storage shed, shop, domestic water building, two well houses and a spring water collection building. There is also a river intake and trapping facility located upstream of Rainbow Lake, along the Tucannon River. There are two residences for staff on site to fulfill security and emergency response needs.

The TFH is supplied with three different water sources. River water is fed from the Tucannon River, and ranges in temperature from 33 to 60 ° F, during use by the hatchery. The intake is located one half mile upstream of the hatchery. This water travels down an open channel into Rainbow Lake. From the outlet of Rainbow Lake the water travels through an 18" above ground pipeline to the hatchery. This pipeline was completely replaced in 2005. Rainbow Lake functions as a reservoir to provide the hatchery with cooler water in the summer months and warmer water in the winter months. It also provides a pool of water to draw from when encountering adverse intake conditions, resulting in temporary loss of water flows. The water right for this source is 16 cfs. Well water is pumped from two separate sources to an aeration tower, and then gravity fed to the rearing units and the domestic pump building. The combined well water right is 2 cfs, with well #2 running around 54 - 57° F and well #3 running a constant 61° F. Spring water is pumped from an underground collection site to the same aeration tower and gravity fed to rearing units. The water right for this source is 5.3 cfs, and has a stable temperature of 51 or 52° F.

The rearing vessels at TFH include forty concrete 1 ft x 15ft x .5 ft shallow troughs, six concrete round ponds approximately 40 ft in diameter with a maximum of 2,660 ft³ of rearing area each, two concrete 10 ft x 80 ft x 3 ft raceways, one concrete 15 ft x 136 ft x 5 ft raceway, and one earthen rearing pond with a maximum of 136,221 ft³ of rearing space. The pond is approximately 170 ft x 200 ft x 6.5 ft in size.

3. Cottonwood Acclimation Facility

Cottonwood AF is located along the Grande Ronde River at RM 28.7, directly above the confluence with Cottonwood Creek in Asotin County, Washington. Construction was completed in February 1985.

This facility includes an adult trapping facility on Cottonwood Creek, and a small storage building. Cottonwood AF has a concrete bottom with earthen walls and holds ~357,000 ft³ of water. It has a water right of 2,694 gpm (6 cfs) for the period January 1st through July 1st. It is supplied with water from Cottonwood Creek through a gravity water supply system, with the intake integrated into the adult trapping facility located ~ 0.10 miles above the pond. Water temperatures range from 34 to 52° F during operation of the facility. It also has a small trailer for use by staff required to be on-site at all times while the pond is in operation. It is presently used for acclimation and release of Wallowa stock summer steelhead into the Grande Ronde River.

4. Dayton Acclimation Facility

Dayton AF is located along the Touchet River at RM 53 in Columbia County, Washington. There is an adult trapping facility on the Touchet River just upstream of the acclimation pond at RM 53.3.

Construction of the Dayton AF was completed in October 1986. This pond is asphalt lined and holds ~ 200,000 ft³ of water. The water right to this pond is 2,694 gpm (6 cfs) for the period of Jan 1st – May 15th of each year. It is supplied with water from the Touchet River through a gravity water supply system, with the intake located at the temporary adult trapping facility just upstream of the pond. Water temperatures during use by hatchery staff for steelhead acclimation range from 34 to 52° F. The pond is located adjacent to the Snake River Lab evaluation office and has a storage garage for equipment and feed. It also has a small trailer for use by staff required to be on-site at all times while the pond is in operation. It is presently used for acclimation and release of LFH stock summer steelhead into the Touchet River.

5. Curl Lake Acclimation Pond

Curl Lake AP is located along the Tucannon River at RM 41 in Columbia County, Washington.

The construction of Curl Lake AP was completed in February 1985. Curl Lake AP is an earthen pond holding ~ 784,000 ft³ of water. It has a water right of 2,694 gpm (6 cfs). It is supplied with water from the Tucannon River through a gravity water supply system. It is currently utilized for acclimation of spring Chinook yearlings for release into the Tucannon River. Water temperatures at this time of year range from 34 to 48 °F. Chinook acclimation in Curl Lake AP started in 1997. After the spring Chinook are released, the pond is stocked with resident trout for fishing. It is emptied after fishing season ends October 31st each year, and recharged by hatchery staff prior to spring Chinook acclimation the following January.

6. Other Acclimation Facilities

In addition to WDFW acclimation sites, LFC provides up to 465,000 yearling and 1,740,000 sub-yearling fall Chinook to three acclimation facilities operated by the Nez Perce Tribe (NPT): Pittsburg Landing and Captain John's Rapids on the Snake River between Asotin and Hells Canyon Dam, and Big Canyon on the Clearwater River. Size at transfer to the NPT AF's is 12 fpp for yearlings and 60 - 75 fpp for sub-yearlings. Size at release goal for acclimated fall Chinook yearlings is 10.0 fpp, and 50 fpp for sub-yearlings.

B. Fish Production Summary

Annual hatchery production is intended to meet LSRCP adult return goals for several species. Current production levels are set to meet the adult return goals for hatchery steelhead most years while minimizing any adverse effects on ESA listed salmon and steelhead (Table 1). Production levels for salmon and steelhead at LFH have been approved through the U.S. v Oregon (US v OR) 2005-2007 Interim Management Agreement. A new production table for fall Chinook (Table 3) was submitted to and approved by U.S. v Oregon parties in 2006 (taken from the draft Snake River Fall Chinook Hatchery Management Plan). LFH is planning fall Chinook production based on this new table. Spring Chinook production includes a conventional and captive broodstock program. With the phase out of the captive broodstock program in 2006, the conventional smolt release program goal will be increased to 225,000 smolts per year (as agreed to under US v OR), for release in 2008. LFH utilizes two steelhead stocks (Lyons Ferry and Wallowa) for mitigation objectives under LSRCP, and is testing two natural broodstocks in the

Touchet and Tucannon Rivers. Numbers of fish released in 2006 and proposed for 2007 and 2008 (Table 2) represent the program as negotiated by the co-managers.

It is important to stress that *any* change to a specific program at LFH or TFH will potentially impact the other programs, so “current capacity” values shown in Table 1 represent rearing limits *as the programs are structured today*. Additionally, restrictions anywhere within the rearing cycle will determine program size. Restrictions can be rearing vessels, water, tagging groups and schedules, fish management decisions regarding harvest or adult return contribution and carrying capacity, etc.

Monitoring and Evaluation (M&E) has been ongoing since 1983 and 1985 for trout and salmon programs respectively. Recent emphasis has centered on meeting Endangered Species Act (ESA) permitting and recovery planning requirements. Routine monitoring includes length, weight, K factor, external fin evaluation, tag retention and fish health examinations. Pre-release quality control checks on fin clips, tag retention, etc. is completed on all WDFW releases by WDFW staff.

Table 1. LFC production capacities (historical design versus current).

Facility	Location River (Mile)	Water Source	Species	Designed Capacity (#Fish)	Designed Capacity (Pounds)	Current Capacity (#Fish)	Current Capacity (Pounds)
Lyons Ferry	Snake (58)	Wells	Fall Chinook	9,160,000	101,800	3,100,000	116,167
			Spring Chinook	132,000	8,800	289,000	9,633
			Steelhead	931,200	116,400	609,500	119,570
			Rainbow	260,000	84,000	310,000	51,600
			TOTALS	10,483,200	311,000	4,308,500	296,970
Tucannon	Tucannon (36)	Wells, Springs, Tucannon R.	Spring Chinook	132,000	8,800	282,000	18,800
			Rainbow	210,000	39,285	198,000	49,100
			Brown (State)	15,000	5,250	-0-	-0-
			Steelhead	-0-	-0-	90,000	20,000
			TOTALS	357,000	53,335	570,000	87,900
Cottonwood AF	Grande Ronde (28.7)	Cottonwood Creek	Steelhead	250,000	31,250	250,000	55,556
Curl Lake AP	Tucannon (41)	Tucannon R.	Steelhead	160,000	32,000	-0-	-0-
			Spring Chinook	-0-	-0-	480,000	32,000
Dayton AF	Touchet (53)	Touchet R.	Steelhead	125,000	25,000	112,500	25,000

Table 2. LFC plants and transfers by brood years (BY) – three-year profile.

Species	Year slated for release/transfer				
	2007 Goal	2007 Actual Plants and Transfers	2008 Goal ^a	Fish/Eggs on Hand For 2008 Goal	2009 Tentative Plan ^b
Fall Chinook					
<u>Yearling releases:</u>	<u>BY 2005</u>	<u>BY 2005</u>	<u>BY 2006</u>	<u>BY 2006</u>	<u>BY 2007</u>
LFH-on station	450,000	503,160	450,000	471,047	450,000
NPT (transfer)	465,000	470,459	465,000	467,233	465,000
<u>Sub-yearling releases:</u>	<u>BY 2006</u>	<u>BY 2006</u>	<u>BY 2007</u>	<u>BY 2007</u>	<u>BY 2008</u>
LFH-on station	200,000	200,692	200,000	unknown	200,000
NPT (transfer)	1,420,000	1,436,712	1,420,000	unknown	1,420,000
Direct- Snake River near Couse Cr (CCD)	200,000	-0-	200,000	unknown	200,000
<u>Fry Transfers</u>					
NPT	-0-	-0-	-0-	unknown	-0-
DNFH/Irrigon-Army Corps of Engineers (ACOE) Transportation Study	-0-	2,000 ^c	-0-	unknown	-0-
<u>Eyed Egg Transfers:</u>	<u>BY 2006</u>	<u>BY 2006</u>	<u>BY 2007</u>	<u>BY 2007</u>	<u>BY 2008</u>
Oxbow - IPC	211,000	127,564	211,000	-0-	211,000
Umatilla - IPC	842,000	-0-	842,000	-0-	842,000
Irrigon - Direct – Grande Ronde R.	421,000	-0-	421,000	unknown	421,000
DNFH/Irrigon-ACOE Transportation Study	328,000	NA	345,220 ^d	-0-	345,220 ^d
Spring Chinook					
	<u>BY 2005</u>	<u>BY 2005</u>	<u>BY 2006</u>	<u>BY 2006</u>	<u>BY 2007</u>
Conventional	132,000	149,466	225,000	108,000	225,000
Captive Brood	100,000	90,056	50,000	82,000	-0-
Summer Steelhead (Stock)					
	<u>BY2006</u>	<u>BY2006</u>	<u>BY2007</u>	<u>BY2007</u>	<u>BY2008</u>
On Station (LFH)	60,000	59,983	60,000	67,600	60,000
Tucannon (LFH)	100,000	96,690	100,000	113,000	100,000
Touchet (LFH)	85,000	86,985	85,000	96,000	85,000
Walla-Walla (LFH)	100,000	97,766	100,000	113,000	100,000
Cottonwood (Wallowa)	160,000	159,242	160,000	179,000	160,000
Tucannon (Endemic)	50,000	62,940	50,000	56,700	50,000
Touchet (Endemic)	50,000	58,989	50,000	68,700	50,000

Spokane Rainbow Trout					
<u>Mitigation</u>					
Catchables	237,500	219,254	236,725	259,234	236,725
Jumbo's	-0-	-0-	500	526	500
Fry-Idaho Fish and Game (IDFG), transfer	160,000	170,125	160,000	NA	160,000
<u>State Program</u>					
Jumbo's	4,000	4,200	4,000	4,500	4,000
Catchables	200	200	200	200	200
Kamloops RB Trout					
Fingerling -IDFG, transfer	50,000	54,343	50,000	NA	50,000

^a all fall Chinook subyearling and egg goals in this column are based on full adoption of the Snake River Fall Chinook Hatchery Management Plan (SRFMP).

^b all fall Chinook goals in this column are based on full adoption of the SRFMP.

^c these fish were transferred to Dworshak National Fish Hatchery (DNFH) at 100 fpp.

^d Amount of fish to transfer to get a 328,000 release. In addition the ACOE has requested that number to be increased to 417,000 (or 438893 at transfer) but an agreement has not been made by US v Oregon parties at this time.

II. SNAKE RIVER FALL CHINOOK

The fall Chinook production program at LFH is the cornerstone of a highly coordinated and integrated artificial program for Snake River fall Chinook, implemented through the LSRCP program, the Idaho Power Company (IPC) Hells Canyon Mitigation Agreement, and the Nez Perce Tribal Hatchery (NPTH). Broodstock for the program at LFH are collected at Lower Granite Dam (LGR) and at LFH, as identified in the 2005-2007 Interim Management Agreement for Upriver Chinook. Co-managers determine the broodstock collection protocol at LGR annually.

The US v OR 2005-2007 Interim Management Agreement determined priority release locations and numbers for fall Chinook production at LFH. As part of development of the Snake River Fall Chinook Hatchery Management Plan, a new fall Chinook prioritized production table was developed (Table 3). This new table was submitted to US v OR parties in 2006, and has been approved, replacing the former table. LFH is planning all fall Chinook production based on this new table.

The LFH was initially designed to release 9.16 million fall Chinook subyearlings (Table 1) at around 90 fpp. Currently this facility produces 1.8 million subyearlings at approximately 50 fpp, and another 900,000 yearlings at 10-12 fpp. Additionally, this facility traps and spawns returning adult fall Chinook to meet egg take needs elsewhere, which includes providing over 1,000,000 eggs annually for the IPC program. It also includes a program change being implemented this year. 421,000 eyed eggs from LFH will be transferred to Oregon Fish and Wildlife (ODFW) for rearing at the Irrigon Hatchery (identified as priorities 13 and 16 in Table 3). Marking and tagging will occur there as well. These fish will be released into the Grande Ronde River in Washington as subyearlings by ODFW. The co-managers will coordinate release timing and location. ODFW fish health staff as coordinated between the two Agencies will

conduct viral testing of the females providing eggs for this program. This production was historically conducted at LFH, however co managers recognized the opportunity to shift the program to Oregon, reducing densities and creating some flexibility at LFH. Both facilities are funded by LSRCP, so budgets were adjusted accordingly, and the co managers have agreed to this change in production.

Table 3. Revised production table listing Snake River fall Chinook salmon production priorities for LFH as documented in the June 1, 2006 Draft of the SRFMP.

Priority	Production Program				
	Rearing Facility	Number	Age	Release Location(s)	Marking
1	Lyons Ferry	450,000	1+	On station	225K AdCWT+VIE 225K CWT +VIE
2	Lyons Ferry	150,000	1+	Pittsburg Landing	70K AdCWT 80K CWT only
3	Lyons Ferry	150,000	1+	Big Canyon	70K AdCWT 80K CWT only
4	Lyons Ferry	150,000	1+	Captain John Rapids	70K AdCWT 80K CWT only
5	Lyons Ferry	200,000	0+	On station	200K AdCWT
6	Lyons Ferry	500,000	0+	Captain John Rapids	100K AdCWT 100K CWT only 300K Unmarked
7	Lyons Ferry	500,000	0+	Big Canyon	100K AdCWT 100K CWT only 300K Unmarked
8	Lyons Ferry	200,000	0+	Pittsburg Landing	100K AdCWT 100K CWT only
9	Oxbow	200,000	0+	Hells Canyon Dam	200K AdCWT
10	Lyons Ferry	200,000	0+	Pittsburg Landing	200K Unmarked
11	Lyons Ferry	200,000	0+	Direct stream evaluation Near Captain John Rapids	200K AdCWT
12	DNFH/Irrigon	250,000	0+	Transportation Study ^a	250K PIT tag only
13	Lyons Ferry ^b	200,000	0+	Grande Ronde River	200K AdCWT
14	DNFH/Irrigon	78,000	0+	Transportation Study ^a	78K PIT tag only
15	Umatilla	200,000	0+	Hells Canyon Dam	200K AdCWT
16	Lyons Ferry ^b	200,000	0+	Grande Ronde River	200K Unmarked
17	Umatilla	600,000	0+	Hells Canyon Dam	600K Ad only
TOTAL	Yearlings	900,000			
	Subyearlings	3,528,000 (of which 328,000 are for Transportation Study)			

^a ACOE Transportation Study wild surrogate groups direct stream released into the Clearwater and mainstream Snake River.

^b for logistical purposes, fish may be potentially reared at Irrigon (LSRCP).

A. Fish on Hand

On September 20, 2007, LFH had an estimated 938,280 (BY06) juvenile Snake River fall Chinook on hand. The program goal is to provide 465,000 yearlings to NPT acclimation sites and 450,000 yearlings for release at LFH in the spring of 2008.

B. Trapping

The trapping goal is 3,426 adults and 198 jacks based upon stray rates and prespawning mortalities encountered in 2006 (Appendix A), and 3,600 eggs per female. This goal is the total number of fish that need to be trapped to meet egg take goals through priority seventeen (Table 3). Generally, between 3,000 and 5,000 fish are trapped. Collection occurs at LFH and LGR. The trapping protocol at LFH is dependent upon what is estimated to be trapped at LGR, the in-season stray rate encountered, and updated return estimates. In effect, trapping is estimated for LGR, and then the remaining numbers of fish needed to meet egg take goals are trapped at LFH. If changes occur in season, the percent trapped at LGR will not change, rather the trapping at LFH changes.

1. Lyons Ferry Hatchery

Trapping at LFH begins in early September, and continues throughout the spawning season, generally ending by late November or early December. All Snake River fall Chinook that voluntarily enter LFH may be retained for spawning. Once the number of fish needed to trap at LFH is estimated, a trapping schedule will be set to reflect the number of fish that need to be trapped weekly, based upon fall Chinook counts at Lower Monumental Dam. When the weekly target is met, no more fish will be retained until the following week. If the hatchery trap is run for steelhead collection and no fall Chinook are needed at the time, the fish will be recycled back to the river. If both fall Chinook and steelhead targets for the week have been met, the ladder will be screened to prevent access to the trap. Since the forecast of fall Chinook returns is less than last year we do not anticipate having to adjust the trapping rate at LFH. All adults trapped at LFH will be retained. Jacks will be trapped and retained not to exceed 15% of the males needed for spawning. Additional jacks may be retained for CWT sampling. Minijacks will be trapped and retained not to exceed 200 fish for CWT sampling.

Coho salmon are occasionally identified at LFH during fall Chinook trapping and spawning operations. WDFW does not propagate coho salmon in the Snake River, but will contact NPT representatives for proper disposition of these fish.

2. Lower Granite Dam

Trapping at LGR may begin as early as August 18 if river water temperatures are less than 70 ° F. Trapping has occurred at a predetermined sampling rate of 11–13% of each hour, twenty-four hours per day. This rate has been increased to 20% for 2007 due to a low pre-season prediction for returns. Collected fish are divided between the LFH and NPTH (usually 70:30) as agreed upon annually, with a predetermined hauling schedule shared between both facilities to meet this need. This hauling schedule is adjusted as appropriate. The trapping/sampling protocol is described more completely in Appendix B.

C. Spawning

Spawning protocols will be consistent with that listed in the draft SRFMP. Spawning will occur weekly, generally on Tuesdays and Wednesdays, starting the third or fourth week in October. It will continue until late November or early December, as necessary to meet egg-take goals. All recovered CWT's will be read or elastomer tags identified during spawning to ensure separation of LFH origin fish from unknown fish. Origin determinations based on scale analysis will be used for untagged fish, and origin based on genetic determination may also be used in 2007.

LFH origin fish (determined by CWT, VI, DNA or scale analysis) will be retained for broodstock. Natural Snake River fish will be incorporated into the broodstock at a target rate of up to 30% (per the SRFMP), provided that this number does not exceed 20% of the natural origin spawning population. Stray (non-LFH origin) hatchery fish as determined by CWT will be culled if not needed by other Columbia Basin hatcheries. Strays based on scale analysis will also be DNA sampled to verify origin. If scales determine the fish is a stray but DNA analysis indicates the fish is an inbasin fish, the DNA results will dictate the final origin of that fish.

Untagged strays may be included in broodstock up to 5%. This limit may be adjusted if necessary to meet production goals and if approved by the co managers. If not needed, strays will be destroyed. Eggs produced from unknown origin brood fish may be utilized in the subyearling program, but not in the yearling program. In addition, Snake River origin (based on WDFW scale readings or genetic samples) unmarked/untagged fish (hatchery and natural), and AD-only fish (from LGR) will be included in the broodstock. It is suggested that unmarked/untagged fish from LGR be used preferentially over unmarked fish at LFH, as they are more likely to be of Snake River origin. This action will be examined on an annual basis. It is the intent of WDFW to exclude out-of-basin fish from the broodstock.

Jacks, fish less than 57cm fork length, will be incorporated into the broodstock at a level not to exceed 15% of the adult males collected. No minijacks (less than 45cm) will be included in the broodstock. Fork length determinations were adjusted based on size at age of CWT fish recovered in 2006.

Our mating protocol is to minimize hatchery stray incorporation into Lyons Ferry Hatchery broodstock while incorporating potentially as many wild fall Chinook as possible. Mating will occur in a 1 x 1 cross. A mating matrix is listed in Appendix C. Because the spawning population is large (>1,000), increasing genetic diversity is not presently a concern. Males may be split and used on multiple females if needed. To maximize the incorporation of wild fish in broodstock, we will scan all untagged fish for PIT tags. If it is determined the fish is wild we will make sure that fish is used in spawning. If the PIT tagged wild fish is a male, we may reserve a portion of the gametes to use on another female.

Fertilized eggs will be water hardened for one hour in 100-ppm iodophore, and incubated in vertical stack incubators. Progeny from below-low enzyme linked immuno-sorption assay (ELISA) females are used for the yearling programs¹. Disposition of eggs from females yielding moderate or high titers during ELISA sampling is determined by co-managers as appropriate. These eggs are used for subyearling programs, or may be culled. Progeny of females not ELISA sampled are only used for subyearlings.

¹ See Section X. for a description of this criterion.

The IPC may receive up to 1,053,000 eyed eggs (842,000 for Umatilla Hatchery + 211,000 for Oxbow Hatchery as part of IPC mitigation), contingent on egg take at LFH and rearing capabilities at IDFG facilities, as was agreed upon in the 1980 settlement agreement.

ODFW’s Irrigon Hatchery may receive up to 421,000 eyed eggs to meet a release goal of 400,000 subyearlings into the Grande Ronde River; contingent on egg take.

There is the potential that surplus Snake River origin adults may be available at the broodstock collection stations once egg take goals have been met. These fish will be returned to the river to continue their upstream migration, or out-planted into natural spawning areas. Adults and jacks released below LGR will be externally marked to ensure they do not compromise run reconstruction efforts at LGR. Table 4 lists the areas that have been identified for each broodstock facility as suitable for disposition of surplus adults.

Table 4. Identified Areas for fall Chinook juvenile and Adult out planting as presented in the June 1, 2006 Draft SRFMP.

Facility	Out plant Locations		
	Adults/jacks	Fry	Subyearlings
Lyons Ferry Hatchery	-Tucannon River -Grande Ronde River -Mainstream Snake River	-Tucannon River -Mainstream Snake River near LFH -Mainstream Snake River above LGR	-Mainstream Snake near Captain John Rapids -Big Canyon -Grande Ronde River -Mainstream Snake downstream of Clearwater River
NPTH	-Lower mainstream Clearwater River -South Fork Clearwater River	-Lower mainstream Clearwater River	-Lower mainstream Clearwater River

D. Rearing

Eggs are reared in the vertical incubators, and are treated with formalin to reduce fungus on a daily basis. They are shocked at eye-up around 550 temperature units (TU’s), and handpicked shortly thereafter. After eggs are picked, folded Vexar sheets are added to each tray for substrate. Formalin treatments stop just before hatch, and after complete yolk-sac absorption by hatched fry (at around 1900 TU’s), they are transferred to raceways for rearing. Head troughs providing well water to the incubators are alarmed, and visual inspections of flow through the trays along with head trough levels are conducted daily.

LFH production fry are moved to outside raceways at ~1,600 fpp. In addition to standard raceways, adult salmon holding raceways are also utilized for subyearling fall Chinook rearing. By utilizing these larger ponds, densities in other raceways are dramatically reduced. Chronic Bacterial Gill Disease has occurred in recent years at LFH and is possibly related to significant

increases in the LFH program. The Bacterial Gill problem is similar to that encountered during the initial years of operation at LFH, when extremely high numbers of subyearlings were programmed. As a result of these density related concerns, the current density index for fall Chinook subyearlings at or smaller than 100 fpp will not exceed 0.08. Density values can increase on a sliding scale to a maximum value of 0.14 for yearlings at 10-12 fpp. These density index goals were developed to improve fish quality and survival.

Yearling fall Chinook are given a 28 day prophylactic treatment using feed treated with erythromycin to reduce the potential for Bacterial Kidney Disease (BKD) outbreaks.

E. Tagging, Transfers, and Releases Assuming Full Production

The following section presumes that the Army Corps of Engineers (ACOE) transportation study will be agreed to for 2008 and lists releases according to the new prioritized production as seen in Table 3. As of 7 November, we anticipate production may be met through priority 15.

A yearling release of 450,000 fish from LFH directly into the Snake River at 10 fpp is programmed. All of these fish will be marked and/or tagged during September (half AD CWT left-red VIE, and half CWT left-red VIE), and transferred into Lake Two. A portion of these fish may also be PIT tagged (as many as 30,000 pending approval) at the same time to better estimate escapement of adults through the hydro system to LFH, LGR, and the Tucannon River (Table 5). Those fish receiving a PIT tag will not be VIE tagged. Fish will be volitionally released from the rearing pond into the Snake River during the period of April 1-15. On or after April 15, all remaining fish in the lake will be forced out, unless river conditions dictate an alternative release time. Since all three lakes share a common release structure, the fall Chinook release must be coordinated with steelhead releases.

Three yearling groups will be marked and/or tagged at LFH in September (AD CWT:CWT only and up to 56,000 PIT tags), then transferred to Captain John, Big Canyon, and Pittsburg Landing acclimation sites (at ~ 12 fpp) for final rearing and release by NPT in April at a target of 10 fpp. Prior to release, NPT staff will PIT tag 5,000 random fish at each site for emigration timing and survival through the hydro-system. This tagging will be coordinated with the COE transportation study. If COE transportation tagging does not occur tagging will be conducted at the acclimation sites.

A total of 200,000 subyearlings are 100% coded-wire tagged and adipose fin clipped in April for release from LFH into the Snake River in early June. As many as 47,222 of this release may be PIT tagged if approval is attained. These fish are pumped directly from the raceways into the river.

Captain John Acclimation Facility receives 500,000 subyearlings in May, as does Big Canyon Acclimation Facility, from LFH. Both groups are comprised of 100,000 CWT, 100,000 AD CWT, and 300,000 unmarked fish. Pittsburg Landing will receive 400,000 subyearlings in May. This group is comprised of 100,000 CWT, 100,000 AD CWT, and 200,000 unmarked fish. All marking and tagging is completed by WDFW in March and April, prior to transfer. Pit tagging may occur prior to and/or post transfer to acclimation sires. These fish are acclimated and released in June by NPT. Prior to release, NPT will PIT tag 2,500 random fish each at Big Canyon and Pittsburg Landing, and 3,500 fish at Captain John, to be compatible with the direct stream released fish out planted at Couse Creek (see next paragraph).

An additional 200,000 subyearlings may be direct stream released into the Snake River at Couse Creek, near Captain John Rapids. These fish are part of a study to compare survival of fish released directly versus those acclimated prior to release. We will coordinate with the NPT to assure that the direct release will correspond with the Captain John acclimated release, scheduled for June. All of these fish will be AD-CWT marked and include 3,500 PIT tags.

ODFW will also direct stream release 400,000 subyearlings into the Grande Ronde River near Cougar Creek (river kilometer 49.4). This group of fish is identified as priorities 13 & 16 (Table 3). They will be transferred to Irrigon Hatchery from LFH as eyed eggs, reared and tagged there, then released into the Grande Ronde River in Washington in early June. 200,000 fish will be AD CWT marked (priority 13), and 200,000 will be unmarked and untagged. WDFW will randomly PIT tag 3,500 fish from this release. The co-managers will coordinate exact release location and timing.

Table 5. Proposed 2007 - 2008 Snake River fall Chinook tagging, transfers and releases.

Site	Goal	Expected at release	Size (fpp)	Age	Mark/CWT/Elastomer	PIT Tags	Transfer/Release Date
Oxbow (IPC)	211,000	Up to 200,000	Eyed Eggs	0+	100% AD CWT	10,000	Jan – Feb 2008 (transfer)
Umatilla (IPC)	842,000	-0-	Eyed Eggs	0+	200K AD CWT 600K AD Only	NA	Jan – Feb 2008 (transfer)
DNFH/research	328,000	-0-	Eyed Eggs	0+	Unknown	328,000	Jan – Feb 2008 (transfer)
LFH	200,000	200,000	50	0+	100% AD CWT	47,222	May – Jun 2008
Grande Ronde Direct - Irrigon	421,000	400,000	Eyed Eggs	0+	200K ADCWT 200K Unmarked	3,500	Jan – Feb 2008 (transfer)
Capt. John	100,000	100,000	50	0+	CWT Only	3,500	Mar – Jun 2008
	100,000	100,000	50	0+	AD CWT		
	300,000	300,000	50	0+	Unmarked		
Big Canyon	100,000	100,000	50	0+	CWT Only	3,500	Mar – Jun 2008
	100,000	100,000	50	0+	AD CWT		
	300,000	300,000	50	0+	Unmarked		
Pittsburg	100,000	100,000	50	0+	CWT Only	3,500	Mar – Jun 2008
	100,000	100,000	50	0+	AD CWT		
	200,000	-0-	50	0+	Unmarked		
Direct near Capt. John	200,000	-0-	50	0+	100% AD CWT	3,500	June 2008
LFH	450,000	450,000	10	1+	225K AD CWT VIE 225K CWT VIE	27,778	April 2008
Capt. John	155,000	150,000	12	1+	70K AD CWT 80K CWT Only	5,000	Feb - 2008 (transfer)
Pittsburg Landing	155,000	150,000	12	1+	70K AD CWT 80K CWT Only	5,000	Mar - 2008 (transfer)
Big Canyon	155,000	150,000	12	1+	70K AD CWT 80K CWT Only	5,000	Mar - 2008 (transfer)

F. Research

The ACOE has made a request for up to 417,000 eyed eggs from LFH for use in an in-river/transportation study, acting as surrogates for natural fish. This request is going through the US v OR process for approval. However, given expected poor adult returns as of September 5, 2007 this request may not be met as identified in Table 3. If eggs are allocated in some fashion for all or a portion of this study, they may be shipped to DNFH or Irrigon Hatchery for incubation and rearing. Alternatively, they may be reared at LFH if space allows. All of the fish would be PIT tagged prior to release, as funded and contracted by the ACOE. Additionally, the National Oceanic and Atmospheric Administration (NOAA) requested 328,000 PIT-tags be divided between all subyearling production releases in the Snake River basin, acting to represent the hatchery component of the in river/transportation study. The LFH portion of these fish will be PIT tagged at LFH, as contracted and funded by the ACOE, and coordinated with hatchery staff.

The co-managers recognize that acclimation prior to release is expected to provide fish performance advantages, however current facility limitations within the basin preclude acclimation of all subyearling groups. A direct versus acclimated study is being conducted by the USFWS, WDFW and NPT to scientifically evaluate the merit of direct stream releases of fall Chinook subyearlings versus acclimated releases. The study will determine if new acclimation facilities in the Snake River basin should be constructed, or are unnecessary. It compares fish performance between groups of the same size (current release size goal is 50 fish/lb), but reared and released under different conditions. Rearing protocols will conform to standard practices, with a focus on maintaining acceptable growth rates, environmental quality, and fish health. Since the managers agree that fish size is critical to the survival of subyearling fall Chinook, size at release will be the primary determinant of release date. Normal acclimation time at CJR is three weeks, and normal release is expected around May 21-25 each year. The acclimation group will be transported to CJR approximately three weeks prior to scheduled release at a projected average size of 75 mm (70 fish/lb). 3,500 PIT tags will be inserted into a random sample of fish within this group prior to release. A second group will be reared at LFH and direct stream released at Couse Creek, just downriver from CJR. They will also have 3,500 randomly inserted PIT tags within this release group. Every effort will be made to meet fish size, and period of acclimation, but the cooperators recognize the potential for early release if fish health will be compromised by environmental or facility conditions. If an early release occurs, the cooperators will coordinate releases as closely as possible.

This study will provide managers with performance comparisons between CJR acclimated and directly released LFH reared subyearling fall Chinook including: (1) passage date at LGR, (2) travel time to LGR, (3) survival from release to the tailrace of LGR, (4) growth and condition measured from release to LGR, (5) smolt-to-adult return rates (SAR's) measured from release to LGR, and (6) spawner fidelity to the Snake River. LGR will be the primary evaluation point for accomplishing all of these objectives with the exception of objective 6.

III. TUCANNON SPRING CHINOOK

The Tucannon River Spring Chinook supplementation program is currently comprised of two broodstock sources, conventional and captive brood. Returning adults trapped at the TFH comprise the conventional broodstock component. A captive broodstock component was initiated in 1997 to prevent extirpation of this listed stock. It was designed to last only one generation (five brood years). This program is being phased out, and 2006 was the last spawn of captive brood adults. The release goal in 2007 was 100,000 yearlings, in 2008 it will be 50,000, and in 2009 no releases will occur.

The conventional program is increasing as the captive program is phased out. The conventional release goal was increased to 225,000 beginning with the 2006 brood year, pending sufficient broodstock.

A. Fish on Hand

On September 20, 2007 LFH had an estimated 107,317 (BY06) conventional juvenile spring Chinook on hand. These fish will be transferred to TFH in October from LFH, and released as yearlings at 9 fpp and 15 fpp from Curl Lake AP into the Tucannon River. Additionally, LFH had 78,759 (BY06) juvenile captive brood spring Chinook on hand. They will be transferred to TFH in October, for eventual release as yearlings from Curl Lake AP at 9 fpp, along with the conventional group.

B. Trapping

Trapping for the conventional broodstock program is conducted exclusively at the TFH adult trap, located just upstream of the hatchery and adjacent to the Rainbow Lake intake. Up to 170 fish (85 wild and 85 hatchery adults – jacks may additionally be collected up to their proportion in the run) will be collected for broodstock, while remaining adults and jacks are counted and released upstream. This increased limit is necessary to meet the new release target of 225,000 yearling smolts. WDFW may collect captive broodstock progeny if run size limits broodstock collection goals, although their use in Broodstock will be limited to as few as possible. WDFW may also retain all of the adult, ESA-listed, Snake River spring/summer Chinook salmon that return to the Tucannon River Fish Hatchery adult trap each year if the total annual adult returns to the trap is less than 105 fish. If the total annual adult returns to the trap are 105 fish or more, WDFW is authorized to retain up to 70 percent of the adult, ESA-listed, Snake River spring/summer Chinook salmon that return to the trap each year and must release at least 30 percent of the adult, ESA-listed, Snake River spring/summer Chinook salmon that return to the trap above the hatchery trap for natural spawning. Adults collected for spawning are transferred by truck to LFH for holding. They are injected in the dorsal sinus at transfer with oxytetracycline and erythromycin, and females only with erythromycin every 30 days until spawning begins. Adults will receive formalin treatments every-other day to control fungus and decrease pre-spawning mortality.

C. Spawning

Conventional broodstock adults will be spawned during September, and the egg goal is 272,000 green eggs. A 2:2 matrix spawning protocol is followed as approved by WDFW Evaluation staff. Fertilized eggs will be water hardened in 100-ppm iodophore for one hour. All spring Chinook carcasses are frozen after spawning, and hauled to the upper Tucannon River for nutrient enhancement, if viral samples test negative.

D. Rearing

Eggs are treated with formalin daily to reduce fungus and reared in vertical incubation trays. At eye-up, they are shocked, handpicked, and substrate is added to each tray. Upon complete yolk-sac absorption (~1600 fpp), they are transferred to outside raceways for introduction to feed and final rearing at LFH.

A prophylactic aquamycin treatment is used to control BKD. This treatment lasts 28 days, and is typically applied in July and August, through feed with 4.5% aquamycin.

In 2005, LFH experienced very high losses in the conventional program during raceway rearing. Hatchery staff believes predation by birds was the primary reason for this loss. In response, the raceways these fish are reared in have been completely enclosed in bird netting. Additionally, other measures have been taken at the hatchery that may contribute to increased survival. Six intermediate fiberglass tanks were purchased and installed, giving culturists more early-rearing space for all programs. This not only reduced densities, it also allows individual spawn groups to be grown together in size before mixing in outside raceways. It also means fish are moved to the raceways at a much larger size, possibly increasing survival to release. Staff also installed an in-line site tube in the venturi vacuum hose, which allows culturists to physically observe the hose to make sure no fish are accidentally vacuumed during routine pond cleaning. Finally, staff have researched various screen seals, and are now using one type for all stocks, proven to be most effective during rearing.

E. Tagging, Transfers, and Releases

In September, the conventional progeny will be 100% CWT/VIE (½ purple and ½ blue non-fluorescent) tagged with no fin clip, and the captive broodstock progeny are 100% coded-wire tagged also with no fin clip (Table 6). Both groups are then transferred to TFH in for final rearing and release. At TFH, both groups are reared in concrete round ponds or raceways on river water, except when well water is added mid-winter to maintain water temperatures above 40° F. Checks for elastomer and CWT retention are conducted prior to transferring the fish to Curl Lake AP in February. For 2008, a release goal of 112,500 conventionals @ 9 fpp & 112,500 conventionals @ 15 fpp (225,000 total) and 50,000 captive brood yearlings at 9 fpp is targeted. All fish will be released from Curl Lake AP in March or April.

Table 6. Proposed 2007 - 2008 Tucannon River spring Chinook tagging, transfers and releases.

Site (Type)	Goal	Expected at release	Size (fpp)	Age	Mark/CWT/Elastomer	PIT Tags	Transfer/Release Date
Curl Lake AP (Conventional)	112,500	52,000	15	1+	100% CWT VIE	2,500	Mar – Apr 2008
Curl Lake AP (Conventional)	112,500	52,000	9	1+	100% CWT VIE	2,500	Mar – Apr 2008
Curl Lake AP (Captive Brood)	50,000	78,000	9	1+	100% CWT	1,000	Mar – Apr 2008

F. Research

In an effort to compare returns based on release size, conventional release numbers will be split in half at marking and reared to two different release sizes. In the spring of 2008 one group will have a target release size of 9 fpp and the other will be at 15 fpp. The final release of the captive brood program fish in the spring of 2008, will be 9 fpp. Studies and practical experience at other facilities suggests a larger release size may increase survival rates.

IV. SUMMER STEELHEAD - GENERAL

The LFC currently uses three stocks of steelhead in the Snake River basin, (LFH, Tucannon, and Wallowa) and two stocks in the Walla-Walla basin (Touchet and LFH). The LFH and Wallowa stocks are both non-endemic stocks that were originally collected from outside their respective release points. The Wallowa stock was originally collected by Oregon Dept of Fish and Wildlife from Lower Snake River dams (likely comprised of both A- and B-run fish from Oregon and Idaho), and then released in the Wallowa River in the Grande Ronde Basin. The LFH stock was derived primarily from a combination of Wells (upper Columbia River) and returning Wallowa stock fish to LFH. The Tucannon and Touchet stocks are both native to their respective streams, though each has had some degree of genetic introgression from the LFH over the years. All of these stocks are collected from a variety of traps located throughout SE Washington (see each stocks description below for specific trapping locations).

The National Marine Fisheries Service’s 1999 Biological Opinion ruled that continued use of LFH and Wallowa steelhead stocks constituted jeopardy to listed steelhead populations in the Snake and Columbia rivers. Concerns about within and out-of-basin straying, and swamping of natural populations by these two hatchery stocks, led NMFS to propose the development of endemic broodstocks where possible, and eventual elimination of non-endemic stocks. Following that ruling, WDFW and the co-managers were responsive to the BIOP by initiating endemic broodstock programs in the Tucannon and Touchet rivers, and have since followed with a decrease in production of the LFH and Wallowa steelhead stocks.

Prior to any of the endemic steelhead being collected for broodstock, WDFW and the co-managers decided that the endemic programs should be tested and evaluated for at least five years at a minimum production level (50,000 smolts annually), before abandoning the LFH stock from production, or increasing the production of endemic stocks. Each endemic broodstock program began with the 2000 BY, with the original goal of collecting 16 pairs for spawning.

Adjustments have been made to the broodstock collections because fecundity and survival values were higher than originally estimated.

The original plan was to utilize adult traps on the Tucannon and Touchet rivers to evaluate the returns and determine success of each program (smolt-to-adult survival rates of the endemic program compared to Lyons Ferry stock releases). However, adult traps have been only partially successful in trapping fish due to high stream flow events. As such, we are now using PIT tags to evaluate each program (smolt-to-adult returns). About 9,000 to 10,000 PIT tags have been into each endemic stock group since 2004. Returns to date indicate that smolt-to-adult survivals of the endemic stock groups have increased (Touchet = 0.4%, Tucannon = 0.7%), but are well below the survivals documented for the Lyons Ferry stock fish releases (1.5%). The smolt-to-adult survival should increase some in the next year or so as rearing modifications at LFH have enabled the endemic stock fish to be released near program size goals (4.5 fish/lb) for the last 2-3 years. Release size goals were generally not met during the first 3-4 years of the program.. Based on the return information to date, WDFW feels there is not enough information available at this time to make an informed decision about stopping the endemic programs or expanding them. An endemic program summary review will be held in early 2008 for all co-managers to review the status of each program.

Adult traps on the Tucannon and Touchet rivers will be used to evaluate the returns and determine success of the program, as well as PIT tag groups in each. Based on the information gathered on returns to date, WDFW feels there is not enough information available at this time to make an informed decision about stopping the endemic programs or expanding them.

V. LYONS FERRY SUMMER STEELHEAD

The LFH stock program was initiated to provide sport fishery opportunities for summer steelhead in the Snake River, it's tributaries, and also includes off-site mitigation in the Walla-Walla Basin. Releases of the LFH stock into the project area have been very successful and adult returns have been reduced in recent years because of ESA concerns.

A. Fish On Hand

On September 20, 2007 LFH had 390,014 (BY07) LFH stock summer steelhead juveniles on hand. These fish will be planted as yearlings into the Snake, Touchet, Tucannon, and Walla-Walla Rivers.

B. Trapping

The LFH stock adults are trapped on-station from volunteers that swim into the fish ladder. The LFH trapping goal is to operate between 1 September and 15 November, which provides adequate adults for the program. Trapping protocols have been set to collect 1,650 fish. Fish are held in large adult holding raceways adjacent to the trap until sorting and spawning. All retained steelhead will be sorted in late November each year. Fish not needed for broodstock or CWT recoveries will be returned to the Snake River for the active sport fishery.

C. Spawning

Spawning will occur in January-February on a weekly basis. Spawning protocol calls for a 2:1 male to female spawner ratio, with each male only being used one time. The intent is to increase the genetic diversity (effective population size N_e) of the hatchery-reared population, and ensure successful fertilization of eggs. Due to IHN virus concerns and expected eggs per female, 120 females will be spawned to produce 530,000 green eggs. Eggs or fry excess to projected program needs will be destroyed or planted as fry in area lakes. All carcasses from spawned fish will be buried on site. All unspawned fish that were retained for broodstock are sacrificed to obtain coded-wire tag or run information.

D. Rearing

After spawning, fertilized eggs are water hardened in 100-ppm iodophore. They are incubated in down-welling iso-incubation buckets (one fish per bucket). After shocking, they are handpicked and weighed down in hatching baskets suspended over shallow troughs. After hatch and swim-up, they are introduced to feed, and transferred to outside raceways at roughly 500 fpp in April. They are reared in these raceways until marking (tagging is completed later) and transferred to Lake One.

E. Tagging, Transfers, and Releases

In August, all LFH stock summer steelhead are adipose fin clipped and transferred to Lake One. In mid-winter, some of these fish are transferred back to raceways to receive additional marks or tags, as determined by WDFW evaluation and Fish Management staff (Table 7). About 87,000 fish are transferred to Dayton AF in mid-February. They are reared there for around 2.5 months, with volitional release into the Touchet River completed by the end of April. In mid-April, 100,000 are trucked to the Walla-Walla River for direct stream release. Also in mid-April, the lower Tucannon River receives 100,000 of these fish by direct stream release. Finally, 60,000 are released from LFH directly into the Snake River in mid-April.

Table 7. Proposed 2007 - 2008 LFH stock summer steelhead tagging, transfers and releases.

Site	Goal	Expected at release	Size (fpp)	Age	Mark/CWT/Elastomer	PIT Tags	Transfer/Release Date
LFH on station release into the Snake River	40,000 20,000	40,000 20,000	4.5	1+	AD Only ADLV CWT	-0- 1,500	April 2008
Dayton AF release into the Touchet River	65,000 20,000	65,000 20,000	4.5	1+	AD Only ADLV CWT	-0- 3,500	Transfer to Dayton AF in Feb, release in April 2007
Direct stream release into the Tucannon River	80,000 20,000	80,000 20,000	4.5	1+	AD Only ADLV CWT	-0- 3,500	April 2008

Direct stream release into the Walla Walla River	80,000 20,000	80,000 20,000	4.5	1+	AD Only ADLV CWT	-0- 3,500	April 2008
--	------------------	------------------	-----	----	---------------------	--------------	------------

F. Research

At this time, there is no direct research associated with the LFH stock summer steelhead at the hatchery (i.e. time or size at release studies, growth studies, etc.). However, starting in 2008, all LFH stock release groups will receive PIT tags (roughly based on proportional release size and expected number of adults returning). Returns from these PIT tags groups will be analyzed separately or as an aggregate to estimate total returns for mitigation accounting purposes. This is partially in response to an anticipated lack of creel personnel in the future to recover CWT's from the summer steelhead fishery.

VI. TOUCHET SUMMER STEELHEAD

The Touchet River summer steelhead is considered an endemic program, meaning all production is derived from natural parentage broodstock. These adults are trapped on the Touchet River at the Dayton AF intake structure and transferred to LFH for holding and spawning. Their progeny are planted in the North Fork of the Touchet River as yearlings each spring.

A. Fish on Hand

On September 20, 2007 LFH had 68,750 (BY07) Touchet River summer steelhead juveniles on hand. These fish will ultimately be direct stream released into the Touchet River at Baileysburg Bridge, roughly 1.5 miles upstream from the Dayton AF.

B. Trapping

Trapping of Touchet River endemic stock begins in January or February (depending on seasonal weather) at the Dayton AF adult trap, located adjacent to the pond intake, and is completed by mid-April. WDFW evaluation staff checks the trap daily, transferring only a portion of unmarked adults to LFH based on broodstock needs. Current survival estimates indicate that 15 spawned females should provide enough eggs to meet the smolt production goal. Therefore, WDFW evaluation staff target collecting 16 females and 20 males for the broodstock (natural origin), with all other wild fish passed upstream for natural spawning. Hatchery fish (LFH stock) are returned to the stream below the trap, to spawn naturally or contribute to the local sport fishery. Hatchery fish (endemic origin) are passed above the trap to spawn naturally in the Touchet River.

C. Spawning

Based on fecundity survival estimates, LFH typically spawns 15 females to provide 73,650 green eggs for the program. Fish in excess to the interim program smolt goals (maximum 75,000 smolts) will be planted into the Touchet River as fingerlings in the fall. Spawning usually occurs in March and April. Matrix spawning is employed, due to the relatively small founding

population for this program. The intent of this protocol is to spawn two males with each female, increasing genetic diversity and successful fertilization of eggs. If not enough males are ripe to achieve this goal; a 1:1 spawning matrix is employed.

D. Rearing

After spawning, fertilized eggs are water hardened in 100-ppm iodophore. They are incubated in down-welling iso-incubation buckets (one fish per bucket). After shocking, they are handpicked and weighed down in hatching baskets suspended over shallow troughs. After hatch and swim-up, they are introduced to feed, and transferred to intermediate raceways at around 500 fpp in June. They are transferred again to outside raceways at roughly 200 fpp in July. In September, they are size-selected during marking and split into two raceways. By sorting into two size groups, culturists can adjust growth rates to minimize size variance at release. Additionally, a number of non-traditional fish culture techniques are being employed on this stock to ensure release size goals are met.

E. Tagging, Transfers, and Releases

In September, all Touchet River endemic stock steelhead are CWT, with no external fin clips (Table 8). They are reared in the raceways until release in April or May at Baileysburg Bridge on the North Fork of the Touchet River. Prior to release, evaluation staff PIT tags 8,000 fish in this group. This will allow for improved data gathering, as these fish are currently not marked for harvest in the sport fishery. The use of PIT tags is an alternate means to calculate smolt-to-adult survivals for program evaluation.

Table 8. Proposed 2007 - 2008 Touchet River summer steelhead tagging, transfers and releases.

Site	Goal	Expected at release	Size (fpp)	Age	Mark/CWT/Elastomer	PIT Tags	Transfer/Release Date
Touchet River	50,000	68,000	4.5	1+	CWT Only	8,000	Apr – May 200

F. Research

At this time, there is no direct research associated with the Touchet River endemic stock summer steelhead at the hatchery (i.e. time or size at release studies, growth studies, etc.). As indicated above, PIT tags along with a CWT's, will give us juvenile migration and SAR data. Other research/monitoring activities are centered on the adult trap (passage issues), and acquiring age composition data from the natural fish.

VII. TUCANNON SUMMER STEELHEAD

The Tucannon River summer steelhead is considered an endemic program, meaning all production is derived from natural parentage. The adults for this program are collected at a temporary trap on the lower Tucannon River, and their progeny planted in the upper Tucannon River as yearlings.

A. Fish on Hand

On September 20 of this year, 56,672 (BY07) Tucannon River summer steelhead juveniles are on hand at LFH. The program goal is 50,000 smolts released. However, because in-hatchery survival of endemic origin fish is unknown, up to 75,000 smolts may be released. If greater than 75,000 smolts are anticipated to be released, up to 25,000 fingerlings could be released into the upper Tucannon River basin in the fall before normal migration.

B. Trapping

Trapping of Tucannon River endemic stock adults begins in September or October at a temporary trap located in the lower Tucannon River (RM 11). Beginning in 2007, the temporary trap may be moved upstream near Marengo (RM 24.5). This is being proposed because we believe many of the wild broodstock being collected are progeny of hatchery origin fish spawning in the lower Tucannon River (near their release location). A trap located further upstream will likely change the time to when broodstock is collected and could have ramifications on spawn timing and rearing at LFH. Current survival estimates indicate that 13 spawned females should provide enough eggs to meet the smolt production goal. Therefore, we will collect 15 females and 21 males (natural origin) for the broodstock at the lower Tucannon Trap, with all other fish (natural, LFH hatchery stock, Tucannon River Endemic Stock) passed upstream for natural spawning. If the lower Tucannon Trap is moved upstream, we propose that all LFH stock fish captured be put downstream of the trap to restrict the upper portion of the basin for just wild and endemic stock origin fish. Some wild fish may also be collected at the TFH adult trap (RM 36) if broodstock collection in the lower river fails. As in the past, all hatchery origin fish (LFH stock) collected at the TFH adult trap will not be passed upstream. Instead they will be marked and released downstream (or taken back downriver below Marengo if the lower trap is moved upstream) to spawn naturally. All endemic and wild fish captured at the TFH will be passed upstream for natural spawning.

C. Spawning

The number of eggs per female is around 6,100. Based on fecundity survival estimates, LFH typically spawns 13 females to provide 79,300 green eggs for the program. Spawning usually occurs in February to early April. Matrix spawning is employed, due to the relatively small founding population for this program. The intent of this protocol is to spawn two males with each female, increasing genetic diversity and helping ensure successful fertilization of eggs. If not enough males are ripe to achieve this goal; a 1:1 spawning matrix is employed.

D. Rearing

After spawning, fertilized eggs are water hardened in 100-ppm iodophore. They are incubated in down-welling iso-incubation buckets (one fish per bucket). After shocking, they are handpicked and weighed down in hatching baskets suspended over shallow troughs. After hatch and swim-up, they are introduced to feed, and transferred to intermediate raceways at around 500 fpp in June. They are transferred again to outside raceways at roughly 200 fpp in July. In September, they are size-selected during marking and split into two raceways. By sorting into two size

groups, culturists can adjust growth rates to minimize size variance at release. Additionally, a number of non-traditional fish culture techniques are being employed on this stock to ensure release size goals are met.

E. Tagging, Transfers, and Releases

In September, all Tucannon River endemic steelhead are CWT tagged, with no external fin clips at LFH (Table 9). In February of 2008, these fish are moved to the TFH. They are reared there until release as yearlings in April or early May. Releases have been roughly five miles upstream of the TFH, at or near Camp Wooten. Prior to release, evaluation staff will PIT tag 8,000 fish in this group. This will allow for improved data gathering because these fish are currently not marked for harvest in the sport fishery. The use of PIT tags is an alternative means to calculate smolt-to-adult survivals for program evaluation.

Table 9. Proposed 2006 - 2007 Tucannon River summer steelhead tagging, transfers and releases.

Site	Goal	Expected at release	Size (fpp)	Age	Mark/CWT/Elastomer	PIT Tags	Transfer/Release Date
Tucannon River	50,000	56,000	4.5	1+	CWT Only	8,000	Apr – May 2008

F. Research

At this time, there is no direct research associated with the Tucannon River endemic stock summer steelhead at the hatchery (i.e. time or size at release studies, growth studies, etc.). As indicated above, PIT tags along with a CWT's, will give us juvenile migration and SAR data. Other research/monitoring activities are centered on the adult trap (passage issues, location of trap), and getting age composition data from the wild fish.

VIII. WALLOWA SUMMER STEELHEAD

The Wallowa stock program was initiated to provide a sport fishery for summer steelhead in the Grande Ronde River (for both Oregon and Washington anglers). It has been an extremely successful program in that regard, and adult returns have warranted a program reduction from a 250,000 yearling release goal to the current program of 160,000 yearlings.

A. Fish on Hand

On September 20, 2007 LFH had 179,711 (BY07) Wallowa stock summer steelhead juveniles on hand. After acclimation at the Cottonwood AF, they are released as yearlings at 4.5 fpp into the Grande Ronde River in April.

B. Trapping

Trapping of returning Wallowa stock adults occurs on Cottonwood Creek (a small tributary to the Grande Ronde River) beginning in March each year. This creek also supplies water to the Cottonwood AF. Trapping occurs from March through April. Because of potential low egg survival and/or IHN virus (both of which have been experienced in the past), about 50 complete spawned females are needed to provide 220,000 green eggs for the program of 160,000 smolts. Unmarked steelhead are not retained for spawning, but passed upstream to spawn naturally. All spawned carcasses will be taken above the trap in Cottonwood Creek and scattered for nutrient enhancement, or returned to LFH to be buried.

C. Spawning

Spawning generally occurs in late March and early April on a weekly basis. All fish are spawned at the Cottonwood Creek trap site, with the gametes transported to LFH for fertilization, incubation, and rearing. Similar to 2007, a 1:1 male to female mating ratio will be employed whenever possible (see research section below). If low water flows in the creek do not allow returning adults access to the trap, two alternate strategies may be employed. First, the acclimation pond outlet creek can be modified to allow adult capture there. Second, excess adults from ODFW's Wallowa Hatchery may be used to provide eggs for this program, as occurred in 2005. Eggs/fry excess to projected program needs will be destroyed or planted in area lakes.

D. Rearing

After spawning, fertilized eggs are water hardened in 100-ppm iodophore. They are incubated in down-welling iso-incubation buckets (one fish per bucket). After shocking, they are handpicked and weighed down in hatching baskets suspended over shallow troughs. After hatch and swim-up, they are introduced to feed, and transferred to outside raceways at roughly 500 fpp in June.

E. Tagging, Transfers, and Releases

In September, these fish are all adipose fin clipped, and 20,000 receive left ventricle clips and are coded wire tagged (Table 10). After marking and tagging, they are transferred to Lake Three at the LFH. In February, they are transferred to the Cottonwood AF for final rearing and release into the Grande Ronde River. 4,000 juveniles will be PIT tagged prior to release in April.

Table 10. Proposed 2007 – 2008 Wallowa stock summer steelhead tagging, transfers and releases.

Site	Goal	Expected at release	Size (fpp)	Age	Mark/CWT/Elastomer	PIT Tags	Transfer/Release Date
Cottonwood AF on the Grande Ronde River	140,000 20,000	140,000 20,000	4.5	1+	AD Only ADLV CWT	-0- 4,000	Transfer to Cottonwood AF in Feb, release in April 2007

F. Research

WDFW has a growing concern that because of the success of the program, and the resultant cuts in production, we have limited the number of fish contributing to the broodstock of the Wallowa program. As such, in 2008, similar to 2006 and 2007, we are conducting a small-scale experiment on the broodstock at Cottonwood Creek to examine the effects of partially spawning females, and then releasing them to continue spawning in the stream. Results from 2006 were very encouraging (23 of 90 kelts were recovered), as 87% of the kelts recovered had less than 10 eggs remaining in the body cavity. The remaining three fish had 100-200 eggs remaining. All fish showed external signs of in-river spawning activity. In 2007, results were similar, but we had a higher incidence of fish that did not spawn after release (~25%). Low stream flows, and a higher incidence of IHNV compromised fish in the broodstock may have contributed to this result. During 2007, 20 redds that were potentially constructed by tagged females were excavated to determine if the eggs were fertilized and developing. Unfortunately, eggs were extracted too soon or too late to positively determine if the embryo was developing.

Identical to 2007, we will partially spawn a maximum of 100 females (compared to the 50 fully spawned females referenced above). Each of the females will be tagged and then released back into Cottonwood Creek above the trap. Evaluation staff and hatchery staff will then periodically survey the stream to 1) identify and mark specific redds made by partially spawned females and 2) to recover any tagged females and assess the degree to which they were able to completely spawn in the creek. Marked redds will be sampled later to determine viability of eggs. With this spawning strategy we will increase and maintain a healthy effective population size for the Wallowa stock program, and gain some valuable insights into the spawning success of females after being partially stripped of their eggs. If documented as successful, this could become a valuable tool for the endemic programs in the Tucannon and Touchet Rivers where founding population size for each of these programs is low, and could be increased to more desirable levels.

IX. SPOKANE AND KAMLOOPS RAINBOW TROUT

Rainbow trout are reared and planted in both southeast Washington and Idaho, to meet LSRCP mitigation goals in both states for lost fishing opportunity as a result of construction and operation of the lower Snake River dams. A small State funded program at the TFH rears rainbow to 1½ pounds each, providing a unique fishing opportunity in local lakes.

A. Fish on Hand

On September 20, 2007 LFH and TFH had 259,234 Spokane stock rainbow trout on hand. LFH also had 54,343 triploid Kamloops stock rainbow trout on hand. These will be shipped to IDF&G in October 2007.

B. Rearing

Eggs for Washington's legal and jumbo programs, along with Idaho's fry plants come from WDFW's Spokane Hatchery (Spokane stock). After receiving these eggs in December and January, a small portion (1,750) is transferred from LFH to regional education programs. Eggs for Idaho's fingerling program are Kamloops stock, from IDFG's Hayspur Hatchery. These eggs are shipped to the TFH in January each year.

180,000 Spokane eyed rainbow eggs for Idaho plants as fry are received at LFH in December. After trough rearing, they are transferred to outside standard raceways in March.

120,000 Spokane eyed rainbow eggs destined as catchables and jumbos are received at LFH in January. Early rearing is conducted in either shallow troughs or intermediate raceways, before transfer to outside standard raceways in April. The following year, they are planted at roughly 3 fpp into local southeast Washington lakes, usually in February and March.

185,000 eyed rainbow eggs (Spokane stock) are received at the TFH in January each year. Of these, 179,000 are destined for planting as legals (3.5 fpp – 137,500 planting goal), and 6,000 are destined for planting as jumbos (1.5 pounds each – 4,000 planting goal). The legal program group is started in shallow troughs, intermediate reared in outside round tanks, and final reared in the earthen rearing pond. The jumbos start in shallow troughs as well, and finish in the round tanks. The entire jumbo program is funded by WDFW.

70,000 Kamloops eyed rainbow eggs are received at the TFH in January. After initial rearing in troughs, they are transferred to outside circular tanks for intermediate rearing. In July at 75 fpp, they are transferred to LFH for marking and final rearing.

C. Tagging, Transfers, and Releases

LFH receives approximately 52,000 Kamloops stock rainbow trout from TFH in July each year, as mentioned above. They are reared in raceways until August or September, when they are adipose fin clipped and either a right or left ventricle fin clipped (alternating years – Table 11). In October, IDFG transports and plants the entire population (usually around 50,000 fish) in Idaho Rivers, at 15 fpp.

No Spokane stock rainbow trout are tagged or fin clipped at LFH. From the raceways, IDFG receives 160,000 fry and transports these fish to designated Idaho waters in April or May, at around 60-80 fpp (Table 12). About 99,225 Spokane stock rainbow trout catchables (@ 2.5 fpp) and 500 jumbos (@ 1 fpp) are planted at 2.5 fpp by LFH drivers into various lakes in southeast Washington. Planting begins in February and is completed in March.

At the TFH, roughly 137,500 Spokane stock rainbow trout are planted into various lakes in southeast Washington as catchables. Planting typically begins in April, and is completed sometime in July. The jumbo trout (usually around 4,000) are planted in February through May each year, supplementing catchable plants.

Table 11. 2007 Kamloops rainbow trout tagging, transfers and releases.

Site	Number	Size (fpp)	Age	Mark/CWT/Elastomer	PIT Tags	Transfer/Release Date
Idaho Rivers	50,000	15	0+	ADLV or ADRV	None	Transfer to and planted by IDFG October 2007

Table 12. 2008 Spokane rainbow trout tagging, transfers and releases.

Site	Number	Size (fpp)	Age	Mark/CWT/Elastomer	Pit Tags	Transfer/release Date
Idaho Reservoirs	160,000	60 – 80	0+	None	None	Transfer to and planted by IDFG in April/May 2008
SE Washington Lakes	236,725 500	2.5 – 4 1	1+ 1+	None None	None None	Planted in February through July 2008
SE Washington Lakes	4,000	1.5 lbs ea	1+	None	None	Planted in February through May 2008

X. FISH HEALTH

A. Guiding Policies

All fish production at LFH is conducted according to the co-managers Salmonid Disease Control Policy and Integrated Hatchery Operations Team (IHOT) fish health policy. Specifically, all lots of fish are monitored for fish health, all broodstock are inspected annually, strict hatchery sanitation procedures and fish culture practices (rearing criteria) are followed, and egg and fish transfer and release requirements are met. BKD (BKD) management strategies for fall Chinook salmon and Infectious Hematopoietic Necrosis (IHN) management strategies for steelhead trout stocks are employed. No management strategy for BKD specific to spring Chinook is currently employed within the LFC.

Currently, IHN in Chinook salmon is not a concern at LFH. The strains of IHN found in the Columbia River Basin have been problematic for sockeye, steelhead and rainbow trout, but not for Chinook salmon. Therefore, standard hatchery practices of egg disinfection and use of pathogen-free rearing water during early rearing have been sufficient fish health measures.

The fish health specialist will respond to all fish disease outbreaks at the request of the fish hatchery staff.

B. Monitoring

The fish health specialist will visit LFH and TFH at least once a month. Mortality records and fish in all rearing containers will be inspected. Approximately 5 - 10 fish of each species may be killed and examined at the discretion of the fish health specialist.

At spawning, all broodstock will be tested for viral pathogens. Ovarian fluid and kidney/spleen samples from at least 60 females will be tested.

C. Specific Fish Health Management

1. BKD Management – Fall Chinook

All female fall Chinook broodstock will receive a pre-spawning injection with erythromycin. All females for use in the yearling production, the IPC program and any others slated for out of state transport will be tested for BKD via ELISA. WDFW categorizes BKD-ELISA optical densities as follows:

- Below-low = < 0.11,
- Low = 0.11 to 0.199,
- Moderate = 0.20 to 0.44,
- High = 0.45 or greater.

Progeny of negative (below low) females will be selected for the yearling fall Chinook program. Eggs from below low and low females will be used for the IPC program. Progeny of all low, moderate and high BKD-ELISA females and untested females may be utilized in the sub-yearling fall Chinook program. These fish will be distributed proportionately among all subyearling releases when possible.

2. IHN Management – Summer Steelhead

All female steelhead broodstock will be tested for IHN virus via cell culture, and the IHN virus levels in the ovarian fluid will be determined.

Eggs from LFH and Wallowa stock females with high levels of IHN virus ($>10^3$) will be destroyed. Eggs from negative and low IHN virus (10^1 to 10^3) females will be reared separately.

Eggs from the Tucannon and Touchet endemic programs with high levels of IHN virus ($>10^3$) may be destroyed, reared separately, or planted into their respective streams as fry, pending agreement among the co-managers. Eggs from negative and low IHN virus (10^1 to 10^3) females will be reared separately.

If IHN outbreaks occur in any fish-rearing vessel, fish from the affected rearing container will be promptly isolated and may be destroyed.

3. Broodstock and Egg Fungus Management

All Chinook broodstock will be treated with formalin every other day or as needed to control fungus. As needed, steelhead broodstock may be treated with formalin. All eggs will be treated with formalin daily to control fungus. Treatment starts 24 hours after the eggs are fertilized. Treatment of steelhead eggs stops when they are weighed into baskets for hatching. Treatment of Chinook eggs stops 7 days before hatching. Rainbow trout eggs are received eyed and are not treated with formalin.

XI. COMMUNICATION

The list of people on the following table (Table 13) are either directly involved in the operation of the LFC, or in related programs and facilities.

Table 13. Contact List.

Name	Agency	Position	Phone	E-mail
Policy Pete Hassemmer Bill Tweit Dave Johnson Gary James	IDFG WDFW NPT CTUIR	Anadromous Coordinator Columbia River Policy Fisheries Dept. Manager Fisheries Program Mgr.	208-334-3791 360-902-2723 208-843-7320 Ext 2442 541-276-4109	phassemmer@idfg.state.id.us twitwmt@dfw.wa.gov davej@nezperce.org garyjmes@ctuir.com
Production Becky Johnson Brian Zimmerman Bruce McLeod Chris Starr Dick Rogers Doug Maxey Harold (Butch) Harty Ron Warren Kent Hills Mike Key Paul Abbott Scott Everett Scott Patterson	NPT CTUIR NPT LSRCP WDFW WDFW NPT WDFW IDFG NPT IPC NPT ODFW	Production Coordinator Production Supervisor Acclimation Facilities Fishery Biologist LFHC Supervisor LFHC Supervisor NPTH Hatchery Manager Hatcheries Division Mgr. Oxbow Hatchery FCAP Hatchery Biologist Coho Recovery Hatchery Coordinator	208-843-7320 Ext 2433 541-966-2376 208-843-7320 Ext 2403 208-378-5329 509-646-3454 509-843-1430 208-843-2825 Ext 2442 360-902-2681 541-785-3459 208-843-7320 Ext 2486 208-388-2353 208-843-7320 Ext 2442 541-963-2138 Ext 22	beckyj@nezperce.org BrianZimmerman@ctuir.com brucem@nezperce.org chris_starr@fws.gov rogerrcr@dfw.wa.gov maxeydwm@dfw.wa.gov hartyhb@nezperce.org warrerrw@dfw.wa.gov oxbowfh@pinetel.com mikek@nezperce.org pabbott@idahopower.com scotte@nezperce.org scott.d.patterson@state.or.us
Evaluation Bill Arnsberg Debbie Milks Jay Hesse Joe Bumgarner Joseph Krakker Mark Schuck Michael Gallinat	NPT WDFW NPT WDFW LSRCP WDFW WDFW	M & E, NPTH Fall Chinook Biologist Research Coordinator Steelhead Biologist Fishery Biologist Evaluations Spring Chinook Biologist	208-476-7296 509-382-1710 208-843-7145 Ext 3552 509-382-1710 208-378-5323 509-382-1004 509-382-4755	billa@nezperce.org milksdjm@dfw.wa.gov jayh@nezperce.org bumqajdb@dfw.wa.gov joe_krakker@fws.gov schucmls@dfw.wa.gov gallimpq@dfw.wa.gov
Management Ed Larson Gary James Glen Mendel John Whalen Scott Marshall Tom Rogers	NPT CTUIR WDFW WDFW LSRCP IDFG	Production Director Fisheries Program Mgr. Fish Management Region 1 Fish Mgmt. LSRCP Coordinator Hatcheries Supervisor	208-843-7320 Ext 2440 541-276-4109 509-382-1005 509-892-7861 Ext 304 208-378-5298 208-334-3791	edl@nezperce.org garyjmes@ctuir.com mendegwm@dfw.wa.gov whalejtw@dfw.wa.gov scott_marshall@fws.gov trogrrs@idfg.state.id.us
Fish Health Kathy Clemens Keith Johnson Sam Onjuka Steve Roberts	USFWS IDFG ODFW WDFW	Supervisory Fish Biologist Fish Pathologist Supervisor Fish Pathologist Fish Health Specialist	208-476-9500 208-939-2413 541-962-3823 509-892-1001 Ext 300	kathy_Clemens@fws.gov kjohnson@idfg.id.us odfwfp@eou.edu robersdr@dfw.wa.gov

Appendix A: 2007 Requests for Production Fish/Eggs (2007Broodyear)

Priority under USvOR (SRFMP) Who	Release site	Age	# for release	transfer	Survival to release or transfer (revised 9/27/07)	Expanded for loss prior release (1/F)	Estim # green eggs to meet priority	SRL Calcs	Total estim eggtake which will cover needs through this priority
1 WDFW	onstation	yearlings	450,000		80.9%	1.23579	556,105	80.9% mean survival, 2004-2000BY	556,105
4NPT	CJ	yearlings	150,000	155,000	80.9%	1.23579	185,368		1,112,210
3NPT	BC	yearlings	150,000	155,000	80.9%	1.23579	185,368		926,841
2NPT	PIT	yearlings	150,000	155,000	80.9%	1.23579	185,368		741,473
			900,000						1,112,210
5 WDFW	onstation	subs	200,000		91.2%	1.09697	219,394	91.2% mean survival, 2004-2000BY	1,331,604
6NPT	CJ	subs	500,000	507,143	91.2%	1.09697	548,486	divided 20K b/t FCAP to acct	1,880,090
7NPT	BC	subs	500,000	507,143	91.2%	1.09697	548,486	for loss from transfer to rel	2,428,576
11 WDFW	direct-Snake R. (CCD)	subs	200,000		91.2%	1.09697	219,394		3,306,483
8NPT	PIT	subs	200,000	202,857	91.2%	1.09697	219,394		2,647,971
10NPT	PIT	subs	200,000	202,857	91.2%	1.09697	219,394		3,087,088
			1,800,000						1,974,550
12 DNFH/Irrigon	Transportation	eyed eggs	250,000	263,125	96.0%	1.04134	274,003	96.0% mean survival, 2005-2001BY	3,580,486
13 WDFW/Irrigon	GRR-direct rel	eyed eggs	200,000	210,500	96.0%	1.04134	219,202	4.99% eye-rel loss	3,799,688
16 WDFW/Irrigon	GRR-direct rel	eyed eggs	200,000	210,500	96.0%	1.04134	219,202	4.99% eye-rel loss	4,323,582
14 DNFH/Irrigon	Transportation	eyed eggs	78,000	82,095	96.0%	1.04134	85,489	4.99% eye-rel loss	3,885,177
9 IPC-Oxbow	HC Dam	eyed eggs	200,000	211,000	96.0%	1.04134	219,723	5.2% eye-rel loss	2,867,694
15 IPC-Umatilla	HC Dam	eyed eggs	200,000	210,500	96.0%	1.04134	219,202	4.99% eye-rel loss	4,104,379
17 IPC-Umatilla	HC Dam	eyed eggs	600,000	631,500	96.0%	1.04134	657,607	4.99% eye-rel loss	4,981,189
			1,728,000						1,894,429

4,428,000 released	4,981,189 green eggs to meet needs through priority 17
number of Snake River origin females needed to spawn	1384 (Estimated using 3600 eggs/F)
Female trapping goal to meet requests through priority 17:	1647 (takes into account strays trapped, prespawn mortalities, nonviable) (LGR+VOL average conversion of .84 for females is used)
	1731 females, spawned 1457 gives .84 conversion
males	1779 (use 90% males, conversion .70 for males LGR + VOL)
jacks	198 (use 10%, conversion .70 for jacks LGR+VOL)
M+F+J to trap:	3624 (LGR+VOL)

Appendix B: 2007 Fall Chinook Trapping/Sampling Protocol

by

Debbie Milks, WDFW
August 17, 2007
Revised September 16, 2007

Executive summary:

The tagging/sampling protocol for broodstock shipped to LFH and NPTH will be the same.

Basics: Scan all FCH for wire and PIT tags. Any fish hauled to LFH or NPTH must be given 1-ROP punch. If you release a fish give it 1-LOP and take scales. Scales should also be taken on 67% of the unmarked/untagged fish hauled to LFH or NPTH.

Note: NO WIRE TAGGED FISH SHOULD BE PASSED UPSTREAM.

Data collected from spring/summer Chinook should be put on the same form that is used for FCH. Please note spring or summer under comments.

Protocol:

- 1) COLLECT & HAUL: All wire tagged FCH adults and jacks (31-52cm). Please give 1-ROP punch.
- 2) COLLECT & HAUL: All unmarked/untagged adult FCH. Please give 1-ROP punch and take scales from two out of every three of these fish.
- 3) PASS: All unmarked/untagged FCH jacks, give 1-LOP punch, and take scales.
- 4) COLLECT & HAUL: All AD Only (no wire) adult FCH, give 1-ROP punch.
- 5) PASS: All AD Only (no wire) jack FCH, give 1-LOP punch, and take scales.
- 6) PASS: All mini-jacks (30cm or less), give 1-LOP punch, and take scales.

More detailed information regarding trapping/sampling:

- 1) Trapping at LGR Dam
 - a. Trapping/Sampling Protocol based upon water temperature in the ladder at the beginning of the day.
 - i. Begin trapping August 18 if temperatures allow
 - ii. Water temps at or below 70° F
 1. Set automatic trapping gates to sample 20% of the entire run, 24 hours a day
 - a. Any fish that are retained for broodstock must receive 1-ROP. If a fish to be retained is accidentally punched on the

left side, give 1-ROP also and make a note in the comments column.

- b. Any fish released must receive 1-LOP and be scale sampled. Place scales in an envelope for age and origin determinations. If these fish are caught again DO NOT scale sample, but enter in data as recapture.

b. Data and Verification

- i. Please note the times you check the trap and when the trap is empty (you are caught up).
- ii. Please write hauling destination (LFH or NPTH) on top of each data form)
- iii. Circle sampling or data recording errors and briefly note in comments column (examples: released with 1-ROP, forgot to scale sample, both sides punched, forgot to record or missing digit in PIT Tag, sample envelope numbers either out of numerical order or skipped for some reason).
- iv. Briefly check over data forms prior to faxing, sometimes erasures and cross-outs are not transmitted clearly through the fax machine.

c. Hauling of broodstock

- i. Injections at LGR Adult Trap
 - 1. All fish collected for broodstock (both LFH and NPTH) will be injected as directed by hatchery staff.
- ii. WDFW and NPT will haul fish from LGR Dam (70% go to LFH and 30% go to NPTH).
 - 1. Fish will be divided weekly unless otherwise agreed to.
 - 2. It was agreed that trucks would be at LGR at 10am when the 70 degree protocol was in effect.

d. Research

- 1. If U of I radio tags fish. Haul all radio tagged fish to LFH or NPT. DO NOT PULL RADIO!!! (we are still waiting to hear about what the Uof I is doing for 2007)
 - a. These fish will be used as broodstock at LFH or NPTH.
- 2. NOAA sort-by-code fish.
 - a. These fish will be used as broodstock at LFH and NPTH.
 - b. Doug Marsh will run a program to indicate which fish were trapped during the 20% and which fish were outside of the trapping period (sort-by-code)

e. Coordination of trapping data and CWT decoding of hauled fish

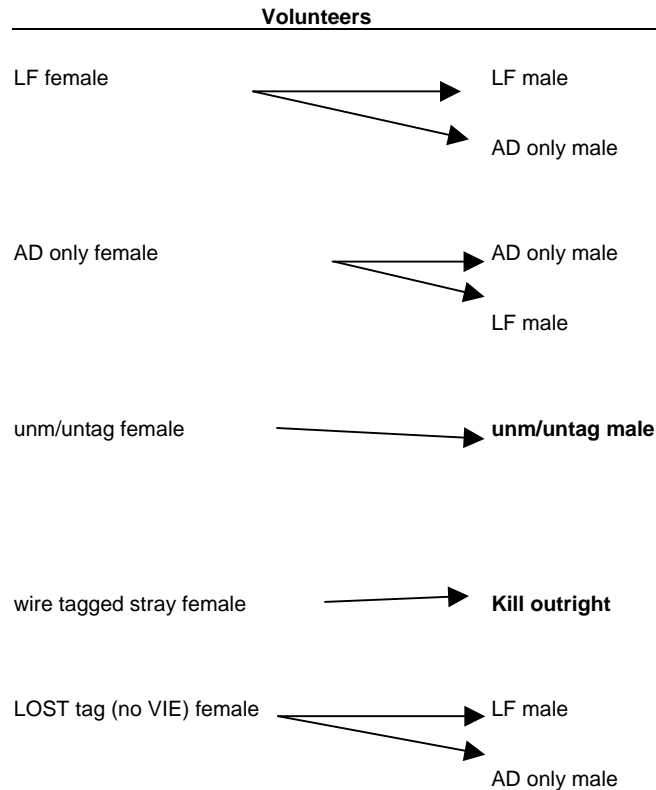
- i. Fax paper copy of data to LFH, NPT, and SRL daily or whenever fish are hauled.
- ii. Data entry, verification, and finalization by January 14.
 - 1. WDFW will enter, verify, and finalize the LGR Adult Trap trapping data.
- iii. All database files at seasons end must be sent to NPT (Bill Arnsberg), WDFW (Debbie Milks), and TAC (Stuart Ellis and Henry Yuen).

f. Video monitoring of sort-by-code fish

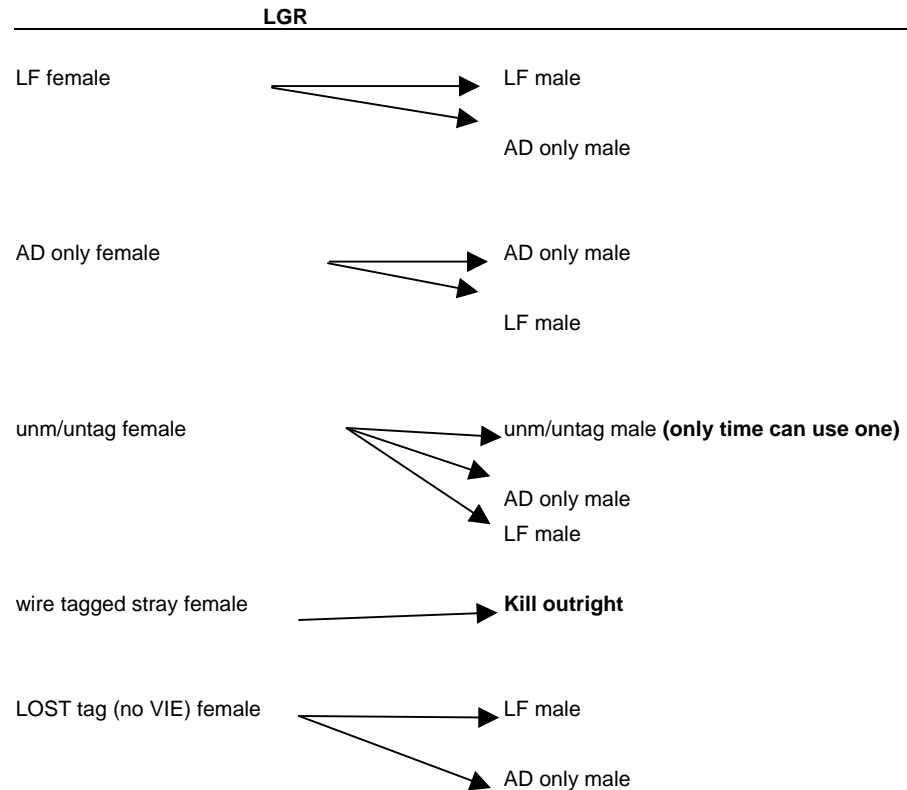
- i. All sort-by-code fish will be video-taped and summarized by Ken McIntyre

A copy of the video and summary will be sent to Debbie Milks so a copy is available for TACs estimating the run comp at LGR

Appendix C: 2007 Mating Matrix for Spawning at LFH (2007Broodyear)



Incorporate jacks in broodstock up to 15%.
Split each LF male (not jacks) and hold to use for LGR day.



Incorporate jacks in broodstock up to 15%.
Reuse Wild origin males (PIT tagged) to maximize progeny with wild genes
Reuse LF origin males as last resort.

CULLING FISH TO REDUCE STRAYS

Culling adults/jacks

wire tagged STRAYS (LGR and LFH trapped)

hold the following gametes separate for possible culling at the end of the season:

- progeny from matings with unm/untagged yearlings (LGR and LFH trapped)
- progeny from matings with unm/untagged STRAY subyearlings (LFH trapped)
- progeny from matings with a parent with unreadable scales (LFH trapped)

