LYONS FERRY COMPLEX ANNUAL OPERATIONS PLAN

For the Period of

OCTOBER 1, 2019 – SEPTEMBER 30, 2020

Prepared by:

Washington Department of Fish and Wildlife



Nez Perce Tribe













Funded By the Bonneville Power Administration through the Lower Snake River Compensation Program



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PREAMBLE

The Annual Operating Plan (AOP) meeting and AOP/SOP documents are planning, coordination and logistics tools that identify the expected implementation of a number of hatchery operation and research/monitoring activities for the coming year in a transparent, open manner.

It is the responsibility of all AOP parties to participate in AOP meetings, provide follow up information and assistance as requested or needed, and work in good faith to complete the AOP document within the timeframe agreed upon at the AOP coordination meeting. A finalized electronic version of the AOP will be available to all cooperating agencies and serve as the working version of the document.

If a disputed or incomplete item is identified at the AOP meeting and persists to the end of the agreed completion timeframe for finalizing the AOP documents, the AOP will be finalized <u>without</u> the disputed or incomplete section. However, parties to the dispute will add a placeholder in the document, so they can work toward resolution.

After the AOP is finalized, and based on unforeseen or unanticipated circumstances (e.g., lower than expected returns, loss of production, infrastructure issues as examples), changes or deviations from the AOP may be warranted. In those cases, there is an expectation that the lead agency that has identified the issue will communicate with the appropriate AOP parties, through the weekly coordination calls or by email, so they can work collaboratively to address it and/or work towards resolution. Implemented changes should be documented in writing by the lead agency and communicated, to ensure transparency and as documentation of the change. These changes should also be captured in various year-end reports.

I. INTRODUCTION

A. Facilities

Lyons Ferry Complex (LFC; See Figure 1) includes Lyons Ferry Hatchery (LFH), Tucannon Hatchery (TFH), Cottonwood Acclimation Facility (Cottonwood AF), Dayton Acclimation Facility (Dayton AF), and Curl Lake Acclimation Facility (Curl Lake AF).

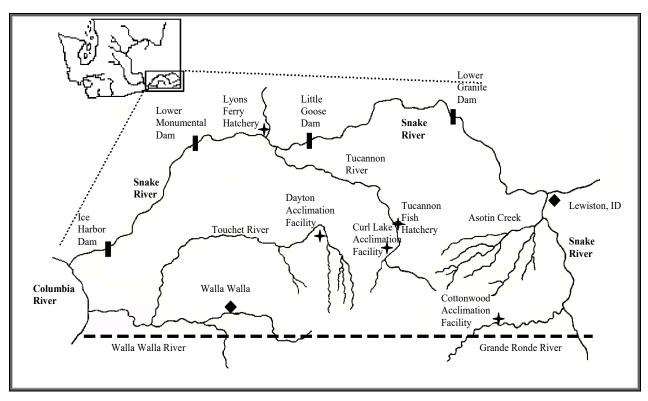


Figure 1. Map of the Lower Snake River Compensation Plan (LSRCP) LFC Facilities, and major rivers and streams in Southeast Washington.

LSRCP funded fish production in Washington began in 1983, with the construction of trout and steelhead rearing facilities at LFH. Construction of salmon hatchery facilities and steelhead acclimation sites followed, and were completed in 1985. Major upgrades at TFH also occurred at that time, and operation of that facility has been funded by LSRCP ever since. Production at all facilities has been directed toward meeting established hatchery adult return goals of 18,300 fall Chinook, 1,152 spring Chinook, 4,656 summer steelhead; plus providing 67,500 angler days of fishing opportunity from 79,000 pounds of rainbow trout production (currently planted at 2.5 fish per pound (fpp). In addition to these LSRCP adult return goals to mitigate for expected hydro system losses (approximately 48% of total desired population returns), the LSRCP hatchery program has contributed to conservation efforts to maintain and restore native populations of salmon and steelhead. Additional hatchery production of jumbo-sized (1.5

pounds each) rainbow trout at TFH that historically was state funded is now funded by the Tri-State Steelheaders (non-profit organization).

1. Lyons Ferry Hatchery

The LFH is located along the Snake River at river mile (RM) 59.1, directly below the confluence of the Palouse River in Franklin County, Washington (Figure 1). Initially it was operated as two separate facilities. Washington Department of Wildlife (WDW) operated the north-side hatchery, producing steelhead and rainbow trout. Washington Department of Fisheries (WDF) operated the south-side hatchery, rearing spring and fall Chinook. A merger of the two agencies in 1994 led to a merging of the two facilities, and has since been operated by the Washington Department of Fish and Wildlife (WDFW) through LSRCP funding as LFH.

LFH facilities include two incubation buildings with office space and feed storage, plus adult fish trapping, holding and spawning structures. A visitor center provides interpretive information for guests of the hatchery. There are eight residences on-site for staff to fulfill security and emergency response needs.

The LFH rearing facilities include twenty-eight raceways at 10 ft x 100 ft x 2.8 ft and nineteen raceways at 10 ft x 88.5 ft x 3.5 ft. The raceways rear all species produced at LFH (spring and fall Chinook, summer steelhead, and rainbow trout). These raceways are covered in 2" square mesh netting. There are three large rearing lakes (643,500 cubic feet (ft³) of water each; 1,100 ft x 90 ft x 6.5 ft dimensions) which are also covered in 2" netting. Netting has significantly reduced predation since being installed in 2006-08. The steelhead and spring Chinook adult holding facilities include three 83 ft x 10 ft x 5 ft adult raceways with an enclosed spawning building incorporated over the center of these ponds. There are four 8.5 ft x 150 ft x 4.3 ft and four 10 ft x 150 ft x 4.3 ft adult fall Chinook salmon holding ponds, which also accommodate fall Chinook subyearling rearing in the spring months. The incubation facilities include 112 full Heath Tray stacks (2 units of 8 trays each) of vertical incubators in the south-side hatching building, and 88 shallow eyeing/hatching troughs and four 3.75 ft x 27.5 ft x 2 ft intermediate rearing troughs in the north-side hatching building.

Water is supplied to LFH from the Marmes pump station, which has emergency power backup generation that was completely upgraded between 2013 and 2016. The Marmes pump (wells) facility has three 300 horsepower (hp) pumps, four 200 hp pumps and one 75 hp pump. The well water right for LFH is 53,200 gallons per minute (gpm), or 118.5 cubic feet per second (cfs) of flow and water temperature is a constant 52°F.

2. Tucannon Hatchery

The TFH is located along the Tucannon River, between the towns of Dayton and Pomeroy Washington, at RM 36 in Columbia County (Figure 1). Fish production began in 1949 by the WDW. In 1983, construction began to remodel the hatchery as part of a transfer of ownership to LSRCP. Since November 1986, when construction was completed, the LSRCP has funded operations.

The TFH includes a combined incubation and office building, back-up power generation building, feed storage shed, shop, domestic water building, two well houses and a spring water collection building. There is also a river intake and adult trapping facility located upstream of Rainbow Lake along the Tucannon River, (Rainbow Lake intake). There are two residences for staff on site to fulfill security and emergency response needs.

The TFH is supplied with three different water sources, (river, well, and spring). River water is captured from the Tucannon River at the Rainbow Lake intake and ranges in temperatures from 33 to 60 °F during use by the hatchery. The Rainbow Lake intake is located one half mile upstream of the hatchery. The captured water from the intake travels down an open channel into Rainbow Lake. From the outlet of Rainbow Lake the water travels through an 18" above ground pipeline (replaced in 2005) to TFH. Rainbow Lake functions as a reservoir to provide the hatchery with cooler water in the summer months and warmer water in the winter months. It also provides a pool of water to draw from when encountering adverse intake conditions along the Tucannon River, resulting in temporary loss of water flows. An estimated 24-36 hours of water supply is currently available following a dredging and restructuring project at Rainbow Lake that was completed in 2018. The water right for the Rainbow Lake intake is 16 cfs. Well water is pumped from two separate sources to an aeration tower, and then gravity fed to the rearing units and the domestic pump building. The combined well water right is 2 cfs, with temperatures from well #2 between 54 - 57 °F and well #3 a constant 61 °F. Spring water is pumped from an underground collection site to the same aeration tower as the well water and gravity fed to the rearing units. The water right for spring water is 5.3 cfs, and has a nearly stable temperature of 51 or 52 °F.

The rearing vessels at TFH include 40 concrete 1 ft x 15 ft x 0.5 ft shallow troughs, six concrete round ponds approximately 40 ft in diameter with a maximum of 2,660 ft³ of rearing area each, two concrete 10 ft x 80 ft x 3 ft raceways, one concrete 15 ft x 136 ft x 5 ft raceway and one earthen rearing pond with a maximum of 136,221 ft³ of rearing space (170 ft x 200 ft x 6.5 ft). Species reared at TFH include rainbow trout, spring Chinook and summer steelhead.

3. Cottonwood Acclimation Facility

Cottonwood AF is located along the Grande Ronde River at RM 28.7, directly above the confluence with Cottonwood Creek in Asotin County, Washington (Figure 1). Construction was completed in February 1985.

This facility includes an adult trapping facility on Cottonwood Creek, and a small storage building. Cottonwood AF has a concrete channel with earthen walls and holds $\sim 357,000~\rm ft^3$ of water. It has a water right of 2,694 gpm (6 cfs) for the period January 1st through July 1st. It is supplied with water from Cottonwood Creek through a gravity water supply system, with the intake integrated into the adult trapping facility located ~ 0.10 miles above the pond. Water temperatures range from 34 to 52 °F during operation of the facility. It also has a small trailer for use by staff required to be on-site at all times while the pond is in operation. It is presently used for acclimation and release of Wallowa stock summer steelhead into the Grande Ronde River.

4. Dayton Acclimation Facility

Dayton AF is located along the Touchet River at RM 53 in Columbia County, Washington (Figure 1). There is an adult trapping and intake facility on the Touchet River just upstream of the acclimation pond at RM 53.3.

Construction of the Dayton AF was completed in October 1986. This pond is asphalt lined and holds ~ 200,000 ft³ of water. The water right to this pond is 2,694 gpm (6 cfs) for the period of Jan 1st – June 1st of each year. It is supplied with water from the Touchet River through a gravity water supply system, with the intake located at the adult trapping and bypass facility approximately 0.3 miles upstream. Water temperatures during operations for steelhead acclimation range from 34 to 52 °F. The pond is located adjacent to the Snake River Lab evaluation office and has a storage garage for equipment and feed. It also has a small trailer for use by staff required to be on-site at all times while the pond is in operation. It is presently used for acclimation and release of both Wallowa and Touchet stock summer steelhead into the Touchet River, and may be used for acclimation and release of Carson stock spring Chinook at some point in the future. The water intake for the Dayton AF, adult trap, and fish ladder structure was rebuilt in 2008 and serves multiple functions. During the late spring and summer months, local irrigators collect water from this intake via a separate screen box and pipeline.

5. Curl Lake Acclimation Facility

Curl Lake AF is located along the Tucannon River at RM 41 in Columbia County, Washington (Figure 1). The construction of Curl Lake AF was completed in February 1985. Curl Lake AF is an earthen pond holding ~ 784,000 ft3 of water. It has a water right of 2,694 gpm (6 cfs). It is supplied with water from the Tucannon River through a gravity water supply system. It is currently utilized for acclimation of Tucannon spring Chinook and Tucannon summer steelhead for release into the Tucannon River. Water temperatures during spring acclimation range from 34 to 48 °F. Chinook acclimation in Curl Lake AP started in 1997 following many years of steelhead acclimation at this site. After the spring Chinook are released in mid-April, steelhead are brought in for a 1-2 week acclimation period and are volitionally released from the time they enter to early May. Following the steelhead release, the pond is stocked with resident trout for fishing. It is emptied after fishing season ends October 31st each year, and recharged by hatchery staff prior to spring Chinook acclimation the following January.

Due to high predation at Curl Lake on spring Chinook pre-smolts, LSCRP provided the funding for a cyclone fence which was installed in early 2015. WDFW staff maintains and monitors the fence and have added an electric wire around the perimeter to keep predators from climbing over the fence. The fence works great for keeping out mammalian predators, but an avian predation issue may still exist. This potentially high predation issue was found through the use of a PIT tag array that was installed at the outlet of the lake which contains five detectors. We will continue to refine the use of the PIT Tag Array and how Curl Lake is drawn down during releases to provide the most accurate estimate of fish released. In 2016, staff also added a propane cannon to scare off avian predators that seemed to help.

6. Fall Chinook Acclimation Project (FCAP)

In addition to WDFW acclimation sites, in 2020 LFC will provide about 1,900,000 subyearling fall Chinook to three acclimation facilities operated by the Nez Perce Tribe (NPT): Pittsburg Landing 600,000 sub yearlings, Captain John's Rapids - 650,000 subyearlings and Big Canyon - 650,000 subyearlings. Pittsburgh Landing will receive two groups of subyearlings, 400,000 in the first group and 200,000 in the second group. Captain John's and Big Canyon will also receive two groups of subyearlings, 450,000 each in the first group and 200,000 each in the second group. Greater details of these facilities and their operations can be found in Appendix D.

B. Fish Production Summary

Annual hatchery production is intended to meet LSRCP adult return goals for several species. Current production levels are set to both conserve and rebuild the Chinook populations, or to meet the adult hatchery return goals for steelhead while minimizing any adverse effects on Endangered Species Act (ESA) listed salmon and steelhead (Table 1). Production levels for salmon and steelhead at LFC have been approved through the U.S. v Oregon (US v OR) 2018-2027 Management Agreement, and specified in species specific tables within. The overall fall Chinook production goal is 4.25 million smolts (includes LFH, FCAP, and Idaho Power Company releases). The spring Chinook production goal is 475,000 smolts per year, 225,000 Tucannon stock for the Tucannon River, (initial release of 225,000 began in 2007 from the original program of 132,000 for the Tucannon stock), and 250,000 Carson stock for the Touchet River (as agreed to under US v OR). LFC is currently utilizing two hatchery steelhead stocks (Wallowa and Tucannon) to fulfill harvest mitigation objectives under LSRCP, and also utilizes two stocks (Touchet and Tucannon), for conservation purposes in the Touchet and Tucannon rivers. The summer steelhead production goal is 585,000 (385,000 Wallowa stock, 150,000 Tucannon Stock, and 50,000 Touchet stock).

It is important to stress that *any* change to a specific program at LFH or TFH will potentially impact the other programs, so "current capacity" values shown in (

Table 1) represent rearing limits as the programs are structured today. Additionally, restrictions anywhere within the rearing cycle will determine program size. Restrictions can be rearing vessels, water, tagging groups and schedules, fish management decisions regarding harvest or adult return contribution and carrying capacity, etc.

Monitoring and Evaluation (M&E) has been ongoing since 1983 and 1985 for trout and salmon programs respectively. Recent emphasis has centered on meeting ESA permitting and recovery planning requirements. Routine monitoring includes length, weight, K factor, external fin evaluation, tag retention and fish health examinations. Pre-release quality control checks on fin clips, tag retention, etc. is completed on all WDFW releases by WDFW staff.

Table 1. LFC production capacities (historical design versus current 2019-20 production goals).

Facility	Location River (Mile)	Water Source	Species	Designed Capacity (#Fish)	Designed Capacity (Pounds)	Current Program Capacity (#Fish)	Current Program Capacity (Pounds)
Lyons Ferry ^a	Snake (59)	Wells	Fall Chinook Spring Chinook Steelhead Rainbow TOTALS	9,160,000 132,000 931,200 260,000 10,483,200	101,800 8,800 116,400 86,000 313,000	3,050,000 475,000 585,000 126,750 4,686,750	131,141 30,033 111,944 48,443 277,357
Tucannon ^b	Tucannon (36)	Wells, Springs, Tucannon R.	Spring Chinook Rainbow Steelhead TOTALS	132,000 210,000 -0- 342,000	8,800 39,285 -0- 48,085	225,000 98,000 50,000 373,000	14,056 43,719 11,111 68,886
Cottonwood AF	Grande Ronde (28.7)	Cottonwood Creek	Steelhead	250,000	31,250	225,000	50,000
Curl Lake AP	Tucannon (41)	Tucannon R.	Steelhead Spring Chinook TOTALS	160,000 160,000	32,000	50,000 225,000 275,000	11,111 18,750 29,861
Dayton AF	Touchet (53)	Touchet R.	Steelhead	125,000	27,750	150,000°	33,333°

^a Lyons Ferry Hatchery was designed to accommodate subyearling Chinook based on the traditional density factor of 0.18. However, with regards to fish health, fish quality, increased yearling production, marking strategies that have been implemented since construction, and water composition, the density index must not exceed 0.09 for subyearlings and 0.14 for yearlings.

^b Tucannon Hatchery was initially designed for rainbow and spring Chinook. Following facility modifications in the 1980's, and the construction of Curl Lake as an acclimation site, increased production for rainbow trout, spring Chinook, and incorporating a steelhead conservation program, were all implemented.

^c 50,000 endemic smolts will be added to the AF when volitional release begins on the Wallowa stock. No feeding will occur at this point and fish will be leaving the pond.

Table 2. LFC plants and transfers by brood years (BY) – three-year profile.

		Year s	Year slated for release/transfer										
Species	2019 Goal	2019 Actual Plants and	2020 Goal ^a	Fish/Eggs on Hand For	2021 Tentative								
		Transfers		2020 Goal	Plan ^a								
Fall Chinook													
Yearling releases: LFH-on station NPT – FCAP (transfer)	BY 2017 450,000 465,000	BY 2017 615,517 281,025	BY 2018 450,000 0	BY 2018 442,742 0	BY 2019 450,000 0								
Subyearling releases: LFH-on station NPT – FCAP NPT – Capt. John 2	BY 2018 700,000 1,900,000 0	BY 2018 725,000 1,971,832 0	BY 2019 700,000 1,900,000 0	BY 2019 0 0 0	BY 2020 700,000 1,900,000 0								
Eyed Egg Transfers: Irrigon-IPC Irrigon - Direct – GRR	BY 2018 1,100,000 ^b 213,000 ^b	BY 2018 1,100,510 213,000	BY 2019 1,100,000 ^b 213,000 ^b	BY 2019 0 0	BY 2020 1,100,000 ^b 213,000 ^b								
Spring Chinook (Stock) Tucannon River (Tucannon) Touchet River (Carson)	BY 2017 225,000 0	BY 2017 152,013 0	BY 2018 225,000 250,000	BY 2018 192,476 260,757	BY 2019 225,000 250,000								
Summer Steelhead (Stock)													
On Station (Wallowa) Touchet (Wallowa) Walla-Walla (Wallowa) Cottonwood (Wallowa) ODFW Wallowa Hat Tucannon (Endemic, Mit) Tucannon (Endemic, Cons) Touchet (Endemic WxW) Touchet (Endemic HxW)	BY2018 60,000 100,000 0 225,000 0 100,000 50,000 25,000 25,000	BY 2018 60,000 100,000 0 225,974 0 118,336 53,579 24,340 15,039	BY2019 60,000 100,000 0 225,000 0 100,000 50,000 0	BY 2019 65,000 110,000 0 230,000 0 64,844 45,617 48,473 0	BY 2020 60,000 100,000 0 225,000 0 100,000 50,000 50,000 0								
Spokane Rainbow Trout													
Mitigation Catchables Jumbo's	BY 2017 197,500 79,000lbs 1,000	BY 2017 215,883° 76,835lbs ^c 1,427	BY 2018 197,500 79,000lbs 1,000	BY 2018 221,749 ^d 79,000lbs 1,200	BY 2019 197,500 79,000lbs 1,000								
IDFG Catchables	1,493lbs 16,000 5,333lbs	2,245lbs 16,200 5,400lbs	1,493lbs 16,000 5,333lbs	1,791lbs 16,200 5,400lbs	1,493lbs 16,000 5,333lbs								
Jumbo's – NPT's	1,650	1,650	1,650	1,650	1,650								
State Program Jumbo's – TSS organization	1,650lbs 4,000 5,714lbs	1,565lbs 4,442 6,439lbs	1,650lbs 4,000 5,714lbs	1,650lbs 4,500 6,428lbs	1,650lbs 4,000 5,714lbs								

^a Based on the *US v. Oregon* table B4.;

^b Transfer numbers include an overage to assure meeting mitigation goals due to possible coagulated yolk .

c Not all fish were released as catchables. 10,015 fish at 5.3fpp & 5,421 fish at 7.8fpp were released in Oct 18.

^d All excess fish will be planted out in Oct 19.

II. SNAKE RIVER FALL CHINOOK

The fall Chinook production program at LFH is the cornerstone of a highly coordinated and integrated artificial program for Snake River fall Chinook, implemented through the LSRCP program, the Idaho Power Company (IPC) Hells Canyon Settlement Agreement, and the Nez Perce Tribal Hatchery (NPTH) with funding through BPA. Broodstock for the program at LFH are primarily collected at Lower Granite Dam (LGR), but may be collected at LFH if trapping at LGR is limited.

The *US v OR* 2018-2027 Management Agreement Table B4 shows priority release locations and numbers for fall Chinook production at LFH. A new production Table has been agreed to by the managers and NOAA Fisheries finished consultation. The new table reflects the most recent changes in production and priorities, and marking/tagging for fish produced from LFH and Irrigon (Table 3).

Table 3. US v OR 2018-2027 Management Agreement Table B4

			Pr	oduction Program				
Priority	Rearing Facility	Number	Age	Release Location(s)	Marking			
1	Lyons Ferry	450,000	1+	On Station	450,000 Ad/CWT			
2	Lyons Ferry	450,000	0+	Captain John Rapids	200,000 Ad/CWT 250,000 Unmarked			
3	Lyons Ferry	450,000	0+	Big Canyon	200,000 Ad/CWT 250,000 Unmarked			
4	Lyons Ferry	500,000	0+	On Station	200,000 Ad/CWT 300,000 Unmarked			
5	Lyons Ferry	400,000	0+	Pittsburgh Landing	200,000 Ad/CWT 200,000 Unmarked			
6	Lyons Ferry	200,000	0+	Captain John Rapids 2	200,000 Ad/CWT			
7	Lyons Ferry	200,000	0+	Big Canyon 2	200,000 Ad/CWT			
8	Lyons Ferry	200,000	0+	Pittsburg Landing 2	200,000 Ad/CWT			
9	Irrigon	1,000,000	0+	Salmon River	200,000 Ad/CWT 800,000 Unmarked			
10	Irrigon	200,000	0+	Grande Ronde	200,000 Ad/CWT			
11	Lyons Ferry	200,000	0+	On Station	200,000 Unmarked			
TOTAL	Yearlings			450,000				
	Subyearlings	3,800,000						

The LFH was initially designed to release 9.16 million fall Chinook subyearlings (Table 1) at around 90 fpp. 2019 production at LFH will be 700,000 subyearlings at approximately 50fpp and another 450,000 yearlings at 10-12 fpp. LFH will transfer another 1,900,000 subyearlings to the FCAP facilities. Size at transfer to the FCAP facilities is 65 - 75 fpp for subyearlings. Size at release goal for acclimated fall Chinook at the FCAP facilities is 50 fpp. Approximately 1,310,000 eyed eggs will be transferred to and reared at the Oregon Department of Fish and Wildlife's (ODFW) Irrigon Hatchery for the LSRCP and IPC programs. The size at release for these subyearling programs are also 50 fpp. 1,000,000 subyearlings will be released into the Salmon River near Hammer Creek for the IPC program and another 200,000 subyearlings will be released into the Grande Ronde River near Cougar Creek as part of LSRCP production. An additional 200,000 subyearlings that were previously reared and released by Irrigon Hatchery into the Grande Ronde River are now reared and released at LFH. This decision was made and agreed upon due to the low success of Grande Ronde River released fish, and the expected greater survival from releasing them at LFH, which will equate to greater downriver fishery contributions, and overall adult returns back to the project area.

A. Fish on Hand

Brood Year 2018

In mid-September 2019, LFH had 442,742 juvenile Snake River fall Chinook on hand. Fish were shipped to Dworshak Hatchery in May and shipped back to LFH the week of August 12^{th} . Marking and tagging began near the end of August. Direct release of 442,700 yearlings at LFH, with ~10,000 of the on-station yearlings PIT tagged at release (Table 4).

Site	Expected Transfer	Expected Release	Size (fpp)	Age	Mark/CWT	PIT Tags	Transfer/Release Date
LFH	N/A	442,700	10	1+	442,700 Ad/ CWT	10,000	Mid-March 2020

Table 4. BY18 Snake River yearling fall Chinook tagging, transfers and proposed releases.

B. Trapping

Brood Year 2019

Tribal, state and federal inter-jurisdictional management of fisheries for conservation of natural populations, sharing of harvestable returns and ESA take, trapping of hatchery broodstocks and distribution of fish trapped in excess of brood needs is extremely complex. In an effort to better coordinate hatchery and harvest management, agencies in the basin have implemented a structured pre-season planning, in-season coordination, post season review and evaluation process. Weekly in-season coordination teleconferences occur and run projections, harvest estimates and hatchery trapping and broodstock collection data are exchanged. Co-managers have agreed to maximize natural origin fish that are incorporated into the broodstock. Trapping protocols at LGR (within reasonable assumptions of what the Lower Granite Trap can handle) and broodstock spawning in-season management will be targeted to achieve 30% proportion of Natural Origin in Broodstock (pNOB) if possible.

The trapping objective, (**Appendix B**), for broodstock at Lyons Ferry is up to 2,600 fish based upon previously observed stray rates and pre-spawning mortalities. The female collection goal for 2019 is 1,323 (\geq 70cm). Each male may be used on multiple females so males are not needed at a 1:1 rate and fewer may be collected. This goal is the total numbers of fish that will need to be trapped to meet egg take goals through *Priority 11*. Brood collection occurs primarily at LGR, but may also occur at LFH or NPTH. Adults trapped at NPTH may be used to supplement LFH production shortages of LGR and volunteer adult returns, and vice versa.

1. Lyons Ferry Hatchery

Trapping at LFH will not occur unless necessary to meet broodstock goals.

2. Lower Granite Dam

Trapping of fall Chinook at LGR is scheduled to begin in early August, but is dependent on water temperature in the trap (Appendix B). In 2019, the trap rate was set at 70% from the beginning of collection through September 6th. On the morning of the 6th, the trap water temperature increased to 71°F. This water was being drawn from 22m below the water surface through the intake flumes that the COE had installed in previous years to get cooler water. Trapping was discontinued the morning of the 6th through the morning of the 12th. By the afternoon of the 12th, NOAA staff were able to start running the trap again as water temperatures were within the operational range. Managers had a few conference calls during this down time and it was suggested and accepted that we would trap at 100% from the afternoon of the 12th through the 16th. Managers had a call on the afternoon of the 16th and suggested dropping the trap rate to 20% through October 6th. The lower trap rate would allow fall Chinook programs to collect enough adults to meet program goals and minimize the excess handling of steelhead. After October 6, IDFG will be able to change the trap rate as needed to further minimize the handling of steelhead if desired. Collected broodstock are divided between the LFH and NPTH (usually 70:30 ratio) as agreed upon annually, with a predetermined hauling schedule shared between both facilities to meet this need, and adjusted as necessary. Additional fish (fish with CWT from an assortment of size and sex) needed for run reconstruction needs will be hauled to LFH. The goal will focus on females in calculating the 70:30 split.

C. Spawning

Brood Year 2019

Spawning will occur weekly, generally on Tuesdays and Wednesdays, starting on the 22nd of October. It will continue until late November or early December, as necessary to meet egg-take goals.

CWT's on males will be read prior to matings to determine origin and age (we want to avoid using strays and jacks if possible). This practice started with BY18 adults. We will continue to increase the percentage of four and five year old fish in the broodstock to offset the past high incorporation rate of jacks in the broodstock and the higher harvest rate of these older age classes in lower river fisheries. Also, the goal for BY19 is to continue the strategy for reducing the

number of "true jacks or jills" (i.e. one-salt fish) in the broodstock. Fork length criteria for broodstock will be adjusted in season to reflect accurate size at age estimates.

Full exclusion of strays in broodstock is preferred to retain Snake River stock integrity. To abide by the US v Or agreement to reach eggtake goals, if broodstock limited, stray females may be included in broodstock as long as matings including a stray do not exceed 5% of the total numbers of matings at LFH. Strays will be incubated separately until we can determine if production goals can be met with Snake River origin females. If the goals can be met without using stays, the progeny will be culled. Jills, (one salt fish), will not be used in production unless it has been determined that we are broodstock limited, but their eggs will be fertilized for fecundity estimates (part of a 5-year evaluation on fecundity). Jills that are spawned are to be mated with true adults. We desire to minimize the numbers of jills in the broodstock so they will be incubated separately until we can determine if production goals can be met with older aged females. If production goals can be met without using jills: 1) the progeny of jills will be culled after fecundity counts at eye up, or 2) released as unfed fry as they would be tagged by Parental Based Tagging (PBT), with option one as the top preference. See Table 5 for disposition of these unfed fry. If we are short on males during spawning, jacks may be used if they come from subvearling production groups. Priority would be to keep known Snake River origin jills before keeping gametes from strays.

The mating protocol, (**Appendix C**), will minimize hatchery stray incorporation into LFH broodstock while incorporating potentially as many wild fall Chinook as possible, striving to maximize pNOB.

Tissue samples (fin clips) will be collected on all broodstock and any unmarked fish during spawning. This action began with the 2011 broodstock. Refer to the Hatchery and Genetic Management Plan (HGMP) and its Addendum for the full intent of the marking and tagging program.

Fertilized eggs will be water hardened for one hour in 100 ppm iodophor and incubated in vertical stack incubators. Distribution of progeny based on BKD ELISA sampling as identified in the fish health section of this document.

There is the potential that surplus Snake River origin fall Chinook may be available at the broodstock collection stations once egg take goals have been met. If so, all LGR transported adults with CWT will be sacrificed and sampled if needed for run reconstruction purposes, and any remaining non-CWT fish will be released back into the Snake River according to Table 5. In the event of broodstock releases during an ongoing fishery, the fish will be marked with a top caudal lobe clip to identify them as fish exposed to MS-222 or Aqui-S 20E. At this time LFH is not using Aqui-S as an anesthetic.

Table 5. Identified areas for fall Chinook juvenile and *adult out planting as presented in the June 1, 2006 Draft SRFMP.

Facility		Out plant Location	s
raciiity	Adults/jacks	Fry	Subyearlings
Lyons Ferry Hatchery	-Tucannon River -Grande Ronde River -Mainstem Snake River	-Tucannon River -Mainstem Snake River near LFH -Mainstem Snake River above LGR -Mouth of Palouse River	-Mainstem Snake near Captain John Rapids -Big Canyon -Grande Ronde River -Mainstem Snake downstream of Clearwater River
NPTH	-Lower mainstem Clearwater River, below North Fork	-Lower mainstem Clearwater River	-Lower mainstem Clearwater River

^{*-}According to fish health guidelines, adults receiving antibiotic injections and/or being anesthetized must meet the withdrawal period for the antibiotic and/or anesthetic used prior to out planting.

D. Rearing

Brood Year 2019

Eggs are reared in the vertical incubators and are treated with formalin at a rate of 1:600 to control fungus on a daily basis. Eggs are shocked at eye-up around 580 temperature units (TU's). After eggs are picked, vexar screening is added to each tray to simulate substrate. Formalin treatments stop just before hatching. Hatched fry are transferred to raceways for rearing after yolk sac absorption at approximately 1,600 fpp, at approximately 1,900 TU's. Head troughs providing well water to the incubators are alarmed and visual inspections of flow through the trays along with head trough levels are conducted daily. BY 2019 will be the second BY in which all release groups will be independently PBT marked, and reared to the release site/time.

In addition to the standard raceways available for rearing fall Chinook, the adult salmon holding raceways are also utilized for subyearling fall Chinook rearing. By utilizing these larger ponds, densities in other raceways are substantially reduced. In addition, this will be the second BY that the on-station subyearling release group will be partially reared (April/May) in one of the large rearing lakes, immediately following the release of yearlings from that same lake. All subyearling release groups will have a 200,000 Ad/CWT group and the remainder of each release group will be unmarked untagged. The 450,000 on-station yearlings will be 100% Ad/CWT, this will be the second year this entire group is Ad clipped. Previously it was 50% CWT only. The current density index for fall Chinook subyearlings up to marking is monitored so as not to exceed 0.09 lbs/ft³/in. Density index values can increase on a sliding scale to a maximum value of 0.14 lbs/ft³/in. for yearlings at 10-12 fpp. These density index goals were developed and agreed upon by all parties to improve fish quality and survival.

E. Tagging, Transfers and Releases

Brood Year 2019

This section outlines the anticipated subyearling and yearling production for BY19 assuming full production of Table 3. All tagging, transfers and releases are listed in Table 6.

Egg Transfers

Irrigon Hatchery will receive 1,310,000 eyed eggs for the IPC program and Grande Ronde direct release (LSRCP program). Eyed eggs are transferred from LFH to the Irrigon Hatchery in mid-December where the fish are reared, marked and tagged prior to release. Coded wire tags for the fish destined for the Grande Ronde will be purchased by WDFW and will have a WDFW Agency prefix. Quality control checks will be completed by WDFW and PIT tags will be inserted by IPC and WDFW staff as part of a cooperative effort. During late May, or by the first week of June, ODFW will direct stream release 200,000 subyearlings at 50 fpp into the Grande Ronde River at Cougar Creek near the Washington border. All 200,000 subyearlings will be Ad/CWT marked/tagged, (*Priority 10* in Table 3), with 4,500 PIT tags. See also Table 6.

The IPC subyearling program at Irrigon Hatchery will receive eggs from LFH in December. Coded wire tags for this release will be funded by IPC and will have an ODFW Agency prefix. Quality control checks will be completed by WDFW and funded by IPC. PIT tags will be inserted by IPC and WDFW staff as part of a cooperative effort. The IPC group is direct released into the Salmon River at the Hammer Creek boat ramp at a release goal of 50 fpp. The release date target is mid to late May. These fish will be 200,000 Ad/CWT and 800,000 unmarked (*Priority 9* in Table 3), with 4,500 PIT tags. See also Table 6.

Sub-Yearling Releases

A total of 700,000 subyearlings with a release goal of 50fpp are intended to be released from LFH into the Snake River in early to mid-May 2020. A total of 200,000 will be Ad/CWT marked and the remaining 500,000 will be unmarked, (*Priorities 4 & 11* in Table 3). See also Table 6. WDFW staff will insert 20,000 PIT tags into the on-station subyearlings as they are being released into the Snake River at the release structure. Quality control checks will be completed by WDFW staff. The 200,000 fish from priority 11 are the last priority of the program and this production will not happen if broodstock needs are not met. The 500,000 fish that will be unmarked will be moved from the raceways directly to Lake 1 once the yearling fall Chinook have been released.

All FCAP release sites will receive two different release groups of subyearlings, (*Priorities 2*, 3,5,6,7 & 8 in Table 3). See also Table 6. Captain John Rapids (CJR) Acclimation Facility (AF) will receive the first group, 450,000, tentatively on April 27, 2020 with a planned release date of May 13. The second group, 200,000, is planned to be received on May 18 with a planned release date of June 4. Big Canyon (BC) AF will receive the first group, 450,000, tentatively on April 27 with a planned release date of May 14. The second group, 200,000, is planned to be received on May 18 with anticipated release date of June 5. Both release sites will have 200,000 Ad/CWT and 250,000 unmarked/untagged fish in the first groups and 100% Ad/CWT fish in the second groups. Pittsburg Landing (PL) AF will receive the first group, 400,000, tentatively on April 20 with a planned release date of May 7. The second group, 200,000, is

planned to be received on May 11 with a planned release date of May 27. The first group will be comprised of 200,000 Ad/CWT and 200,000 unmarked/untagged fish and the second group will be 100% Ad/CWT.

All marking and CWT tagging is completed by WDFW in March and April, prior to transfer. PIT tagging may occur prior to and/or post transfer to acclimation sites. All of these subyearling groups are acclimated and released by NPT at a goal of 50 fpp. Quality control checks, PIT tagging, and the purchase of the PIT tags for fish destined for FCAP facilities will be completed by NPT staff.

Yearlings

A yearling release of 450,000 fish from LFH directly into the Snake River at 10 fpp is programmed for 2021. All of these fish will be Ad/CWT'd during August-September 2020 and transferred into Lake 1. A portion of these fish will also be PIT tagged at release. In 2021, these fish will be released over an anticipated 3 to 4 day period into the Snake River in mid- to late-March, depending on river flows.

Table 6. Proposed BY19 Snake River fall Chinook tagging, transfers and releases.

Site	Transfer	Release	Size	Ag	Mark/CWT	PIT	Transfer/Release
	Goal	Goal	(fpp)	e		Tags	Date
Irrigon (IPC)	1,100,000	1,000,000	Eyed	0+	200,000 Ad/CWT	4,5001	Dec 2019 (eggs transfer)
			Eggs		800,000 Unmarked		
Grande Ronde	210,000	200,000	Eyed	0+	200,000 Ad/CWT	4,5001	Dec 2019 (egg transfer)
Direct - Irrigon			Eggs				,
LFH	N/A	700,000	50	0+	200,000 Ad/CWT	$20,000^{1}$	May 2020
		,			500,000 Unmarked	.,	
Capt. John 1	451,000	450,000	75	0+	200,000 Ad/CWT	$26,000^2$	April – 2020 (transfer)
					250,000 Unmarked		
Big Canyon 1	451,000	450,000	75	0+	200,000 Ad/CWT	$11,000^2$	April - 2020 (transfer)
					250,000 Unmarked		, , ,
Pittsburg	401,000	400,000	75	0+	200,000 Ad/CWT	$26,000^2$	April – 2020 (transfer)
Landing 1					200,000 Unmarked		
Capt. John 2	201,000	200,000	75	0+	200,000 Ad/CWT	$4,500^3$	May - 2020 (transfer)
1							, , ,
Pittsburg	201,000	200,000	75	0+	200,000 Ad/CWT	$4,500^3$	May – 2020 (transfer)
Landing 2							
Big Canyon 2	201,000	200,000	75	0+	200,000 Ad/CWT	$4,500^3$	May - 2020 (transfer)
							, , ,
LFH	N/A	450,000	10	1+	450,000 Ad/CWT	$10,000^{1}$	March 2021

^{1.} Provided by LSRCP

² Provided by the Fish Passage Center for Comparative Survival Studies (CSS).

^{3.} Provided by the Bonneville Power Administration (BPA).

III. TUCANNON SPRING CHINOOK

The Tucannon River Spring Chinook Hatchery production began in 1985 using endemic broodstock. Currently, both natural origin and hatchery supplementation fish are collected for broodstock. Returning adults are collected at the Rainbow Lake Intake and transported to LFH for holding, spawning, hatching and initial rearing. The release goal is 225,000 yearling smolts. WDFW has initiated internal discussions on different rearing/release strategies for the future.

A. Fish on Hand

Brood Year 2018

Mid-September 2019, LFH had 192,746 juvenile spring Chinook on hand. Fish were shipped to Dworshak Hatchery in May and shipped back to LFH the week of August 12th. Approximately 6,000 fish have been segregated due to high Elisa results for BKD at Lyons Ferry. This group was moved into Lake 2 at LFH the second week of September. This is the first year this stock has been reared in one of the lakes at LFH. They will then be shipped directly to Curl Lake for release in early/mid-March and not overwintered at the Tucannon Hatchery as done in the past.

B. Tagging, Transfers, and Releases

Brood Year 2018

In March 2019, the BY18 progeny were 100% CWT tagged with no fin clip at LFH.

The BY18 spring Chinook at LFH will be reared in a lake at LFH for the first time. Overwintering at TFH will not happen as the fish will go directly from LFH to Curl Lake in early/mid-March for a planned two week acclimation. Checks for CWT retention are conducted prior to transferring the fish to Curl Lake AF in March when the fish are PIT tagged from the release structure at LFH. PIT tagging will be performed by evaluation staff in 2020 instead of by Biomark Inc. as has been done in previous years.

Table 7. Proposed BY18 Tucannon River spring Chinook tagging, transfers and releases.

Site (Type)	BY18 Release Goal	Expected at release	Size (fpp)	Age	Mark/CWT	PIT Tags	Release Date
Curl Lake AP	225,000	192,000	12	1+	100% CWT	15,000	March 2020

C. Spawning / Outplants

Brood Year 2019

The egg take goal for BY19 is approximately 245,000 green eggs. Seventy-five females are needed to meet the egg take goal at a fecundity of 3,500. For BY19, 44 females were spawned with an estimated fecundity of 2,800. This is approximately 120,000 green eggs. Low spawner numbers were due to a high prespawn mortality at TFH and then continued at LFH after the fish were transferred back to LFH in early August. Due to the pipeline leak at LFH, all adults were initially held at TFH when collected. Fish being held for broodstock were inoculated for a second time on August 5th as they were loaded onto the trucks to go to LFH.

A 2 x 2 spawning matrix protocol will be followed for spring Chinook spawning at LFH. During the spawning activity, eggs and milt are collected in individual bags and placed in a cooler until fertilization. Spawning matrices are determined after all fish are spawned, all CWT's are checked for origin, and then fertilization takes place at the spawning building. Fertilized eggs are then brought to the dirty room where they are laid down individually into heath baskets, rinsed and placed into heath stacks to water harden in 100 ppm iodophor for one hour. All prespawn mortalities and spawned spring Chinook carcasses are disposed of on site, or will be used for nutrient enhancement in the upper Tucannon River if possible.

Due to high pre-spawn mortality of adults passed above the trap in the Tucannon River in the past, and expected low returns for 2019, agreement was reached to bring all adults that would be passed above the trap back to LFH to be held for spawning or adult out planting. WDFW has been doing this for four years now. There were no adults for out planting for BY19, but some jacks (~20) were outplanted above the Tucannon FH intake as some adults were observed during pre-spawn walks in the upper basin.

D. Rearing

Brood Year 2019

The production goal for BY19 is 225,000 smolts at release (Table 8), but will not be achieved due to the high pre-spawn loss experienced this year. Eggs are treated with formalin daily to reduce fungus and are reared in vertical incubation trays. At eye-up, eggs from individual females are shocked, picked and placed in separate trays with vexar screening to simulate substrate. Upon complete yolk-sac absorption (~1600 fpp), they will be transferred to the north side shallow troughs for introduction to feed or ponded directly into raceways on the south side of LFH.

Table 8. Proposed BY19 Tucannon River spring Chinook tagging, transfers and releases.

Site (Type)	BY19 Release Goal	Expected at release	Size (fpp)	Age	Mark/CWT	PIT Tags	Transfer/Release Date
Curl Lake AP	225,000	110,000	12	1+	100% CWT	15,000	March 2021

E. Trapping

Brood Year 2020

Trapping for the spring Chinook broodstock program is conducted exclusively at the TFH adult trap, located upstream of TFH and incorporated to the Rainbow Lake Intake. Broodstock collection is permitted up to 170 adults. The proportion of hatchery and natural origin adults incorporated into the broodstock is based on the estimated run size and the Tucannon Spring Chinook HGMP sliding scale (Appendix H) and will be adjusted in-season, if necessary, to meet the 225,000 smolt production goal. One-ocean age (jacks: <61 cm FL) fish may be included in the brood at a rate not to exceed 10% of the adult males during low run years.

Adults collected for spawning are transferred by truck to LFH for holding. Adults will receive 167 ppm formalin treatments every-other day to control fungus and decrease pre-spawning mortality

Depending on the pre-season forecast, experience from past out planting success, and expected environmental conditions in the Tucannon River in 2020, WDFW (with co-manager agreement) may collect and hold some portion or all of the returning adults that would normally be passed upstream to spawn naturally. All held fish will be brought back to LFH and released back into the river just prior to spawning in August 2020. Percentage collected will be dependent upon preseason run forecasts and actual numbers back to the river. Conversations will be ongoing up to and through adult collection.

Staff will pass, or collect for holding, hatchery jacks to mimic the NOR jacks returning to the best of their ability and cull the excess hatchery jacks at the trap. Jacks culled at the adult trap will be utilized for food bank or stream enrichment purposes. On a low male proportion year, we will pass more HOR jacks to help ensure there are enough males on the spawning grounds.

IV. ASOTIN CREEK SPRING CHINOOK

WDFW has begun exploring options with the co-managers to implement a spring Chinook program in Asotin Creek using Tucannon River stock. This program would allow for another broodstock source in years of low returns, and would act as a safety net to the Tucannon River population. Much planning still needs to occur, along with ESA consultation with NOAA Fisheries before such a program can be implemented. WDFW has completed a draft HGMP for this program at the currently agreed to program level of 75,000 smolts. WDFW is committed to work with the co-managers and LSRCP over the coming year to develop an agreed upon plan, and finalize up the HGMP.

V. TOUCHET SPRING CHINOOK

WDFW brought forth a proposal to PAC in January of 2018 to initiate a harvest mitigation program for spring Chinook in the Touchet River. This proposal was agreed to in PAC, passed on and accepted through the US vs OR Policy Committee. The HGMP for this program has been submitted to NOAA Fisheries for consultation. WDFW received ~275,000 eyed Carson stock eggs from the USFWS Little White Salmon facility in 2018 to produce 250,000 smolts to be released in the spring of 2020. Hatching and rearing has taken place at LFH. Fry will be ponded and reared in raceways until they are shipped to the Dayton Acclimation Facility in January/February 2020 (see table 9). Smolts will be PIT tagged before they are moved to the Dayton AF. Smolts will be released from the Dayton AF in March 2020. Descriptions of future broodstock collections, spawning and rearing of this group of fish at LFH will be provided in future AOP's.

A. Fish on Hand

Brood Year 2018

Mid-September 2019, LFH had 260,757 juvenile spring Chinook on hand. Fish were transferred to Dworshak hatchery near the end of May to facilitate the repair of the intake line coming from the Marmes pump station to LFH. Fish were moved back to LFH the week of August 12th.

B. Tagging, Transfers, and Releases

Brood Year 2018

In March 2019, the BY18 progeny were 100% Ad clipped and CWT tagged at LFH.

Table 9. Proposed BY18 Tucannon River spring Chinook tagging, transfers and releases.

Site (Type)	BY18 Release Goal	Expected at release	Size (fpp)	Age	Mark/CWT	PIT Tags	Release Date
Dayton AP Facility	250,000	260,000	12	1+	85,000 Ad/CWT 165,000 Ad Only	15,000	March 2020

C. Spawning

Brood Year 2019

The goal for eyed eggs to be shipped in is 270,000. For 2019, eyed eggs are to be provided from Little White Salmon NFH.

D. Rearing

Brood Year 2019

The production goal for BY19 is 250,000 smolts at release (Table 10). Eggs are treated with formalin daily to reduce fungus and are reared in vertical incubation trays. At eye-up (if we receive green eggs), eggs from individual females are shocked, picked and placed in separate trays with vexar screening to simulate substrate. Upon complete yolk-sac absorption (~1600 fpp), they will be ponded directly into raceways at LFH.

Table 10. Proposed BY19 Touchet River spring Chinook tagging, transfers and releases.

Site (Type)	BY19 Release Goal	Expected at release	Size (fpp)	Age	Mark/CWT	PIT Tags	Transfer/Release Date
Touchet River	250,000	250,000	12	1+	85,000 Ad/CWT 165,000 Ad Only	15,000	March 2021

E. Trapping

Brood Year 2020

For BY20 there is no plan of trapping adults at the Touchet River trap. We will not see any returning adults until 2022. At that time WDFW will begin trapping broodstock but will also receive eggs from Carson or Little White Salmon NFH's until we have enough adults returning to meet broodstock needs.

VI. SUMMER STEELHEAD - GENERAL

The LFC currently uses two stocks of steelhead in the Snake River basin, (Tucannon and Wallowa) and two stocks in the Walla Walla basin (Touchet and Wallowa). The Wallowa stock is a non-endemic stock and was originally collected by ODFW from Lower Snake River dams (likely comprised of both A- and B-run fish from Washington, Oregon and Idaho), and then released in the Wallowa River in the Grande Ronde Basin. The Wallowa stock steelhead will be released in the Grand Ronde and Touchet rivers, and on-station at LFH into the Snake River.

The NMFS 1999 Biological Opinion ruled that continued use of Lyons Ferry and Wallowa stocks were causing jeopardy to listed ESU Steelhead populations in the Snake and mid-Columbia rivers. It was recommended by NMFS to convert to endemic stock populations where possible. The Touchet and Tucannon endemic broodstock programs began with BY2000.

Additional changes to the steelhead program are likely in response to results from evaluation of fish stock performance and ESA related concerns regarding the ongoing releases of Wallowa stock steelhead into the Snake, Touchet and Grande Ronde rivers. Such changes may require a departure from the general mitigation approach used for steelhead so far, but also will need careful planning to ensure that the change can be implemented within the limits of the hatchery facilities now or as planned to exist in the near future.

For BY 2019, steelhead were held at three facilities to allow for the main pipeline at LFH to be repaired. Wallowa stock eggs were taken at the ODFW Wallowa Hatchery and then shipped to Irrigon hatchery as eyed eggs. They were early reared at Irrigon and then shipped back to LFH the week of August 5th after the pipeline was repaired. Tucannon stock were shipped to TFH as eyed eggs and then shipped back to LFH the week of August 5th. Touchet stock were held in the trough room at LFH and water was supplied by pumps other than the main pumps at Marmes.

VII. TOUCHET SUMMER STEELHEAD

The Touchet River summer steelhead program is considered an endemic program. Through BY2014, all production was derived from natural parentage broodstock. Between BY2015 to BY2018, per co-manager agreement, WDFW conducted a study which incorporated hatchery origin returns (Touchet stock – CWT only fish) into the broodstock to test smolt to adult survival differences, and whether incorporating hatchery fish in to the program would increase the relatively poor survival we've observed from this program since it begin in 2000. During fertilization, crosses consisted of either WxW or HxW origin fish. To simplify the matings and during broodstock collection females were either hatchery or wild origin, while only wild origin males were collected each year. For each study year, 5,000 PIT tags each were inserted into each rearing group (WxW or HxW). See the research section from preliminary results to date. Adult returns from the study will be complete in the 2021 run year, and a management decision about how to move forward with this program will occur at that time.

With the initial part of the study complete (broodstock crosses and rearing), broodstock collections switched back to 100% wild origin fish for broodstock in 2019. This is consistent with the HGMP that was submitted and approved by NOAA Fisheries while the matings study results are pending.

Broodstock adults are trapped on the Touchet River at the Dayton AF intake structure and transferred to LFH for holding and spawning. Historically progeny from this program were planted in the North Fork of the Touchet River as yearlings each spring. Starting in BY15 (same year the mating study was initiated), smolts were trucked to the Dayton AF and allowed 10 to 14 days to acclimate with the Wallowa stock at the time the volitional release began. At the end of the acclimation period, the remaining fish will be forced out to the Touchet River. All adults trapped and handled are anesthetized by electronarcosis (EN).

A. Fish on Hand

Brood Year 2019

Mid-September 2019, LFH had approximately 48,473 Touchet River summer steelhead juveniles on hand.

B. Tagging, Transfers, and Releases

Brood Year 2019

In September and October, all Touchet River endemic stock steelhead were CWT tagged, with no external fin clips, thereby making them unsusceptible to sport fisheries. Smolts will be put into the Dayton AF and allowed to comingle with the Wallowa stock and then volitionally outmigrate with the Wallowa stock.

Table 11. Proposed BY19 Touchet summer steelhead tagging, transfers and releases.

Site	BY18 Goal	Expected at release	Size (fpp)	Age	Mark/CWT	PIT Tags	Transfer/Release Date
WxW - Touchet River (Dayton AF)	50,000	47,000	4.5	1+	100% CWT	5,000	April 2020

C. Trapping

Brood Year 2020

Trapping of BY20 Touchet River endemic stock will begin in January or February (depending on seasonal weather) at the Dayton AF adult trap (located adjacent to the pond intake) and is generally completed by early May. WDFW evaluation staff checks the trap daily, using EN to calm the fish for handling, transferring only a portion of natural origin adults to LFH based on broodstock needs. All remaining NOR's and any captured endemics are released upstream of the trap. All trapped Wallowa stock fish are: 1) transferred to the Dayton Juvenile Fishing Pond to remove them from the river and provide additional fishing opportunities within Dayton, 2) sacrificed for CWT retrieval, and/or 3) donated to a local food bank.

Current survival estimates indicate that 14-15 spawned females (depending on age structure) should provide enough eggs to meet the smolt production goal (Table 11). Per co-manager agreement, WDFW evaluation staff target collecting 16 females and 20 males for the broodstock (100% natural origin)

D. Spawning

Based on fecundity and survival estimates, LFH typically spawns 14-15 females to provide 65,000 green eggs for the program. Up to 60,000 smolts may be reared full cycle and planted as yearlings in the spring. Fish in excess of 60,000, will be planted into the Touchet River as fingerlings in the fall, untagged. Spawning usually occurs in March and April. A matrix-type spawning protocol is employed, (2x1; two males to every female), to increase the effective breeder population (N_b) due to the relatively small founding population for this program. If not enough males are ripe to achieve this goal; 1:1 spawning is employed.

E. Rearing

After spawning, fertilized eggs are water hardened in 100 ppm iodophor. They are incubated in down-welling iso-incubation buckets (one fish per bucket) until the eggs eye up. The eyed eggs are shocked, ran through an automated egg sorting machine or handpicked and enumerated, and placed in hatching baskets suspended over shallow troughs. After hatch and swim-up, they are introduced to feed, and transferred to intermediate raceways at around 500 fpp in June. They are transferred again to outside raceways at roughly 200 fpp in July.

Table 12. Proposed BY20 Touchet summer steelhead tagging, transfers and releases.

Site	BY20 Goal	Expected at release	Size (fpp)	Age	Mark/CWT	PIT Tags	Transfer/Release Date
WxW - Touchet River (Dayton AF)	50,000	50,000	4.5	1+	100% CWT	5,000	April 2021

VIII. TUCANNON SUMMER STEELHEAD

The Tucannon River summer steelhead program is considered an endemic program, meaning all original production was derived from natural parentage, and in later years, from 1st generation hatchery reared endemic stock fish as well. The adults for this program are collected at TFH and their progeny planted in the Tucannon River as yearlings. Current release goal is 150,000 smolts at 4.5 fpp, with 50,000 smolts being released for the conservation portion of the program (unclipped) and 100,000 smolts being released for the mitigation portion (ad-clipped) of the program. According to the Tucannon Steelhead Program broodstock sliding scale, the 50,000 smolts for the conservation portion will come from NOR's and unclipped endemic returns (conservation group). The 100,000 smolts for the mitigation portion will come from hatchery endemic returns and consist of ad-clipped/cwt or cwt-only adults, with no NOR's at lower NOR return levels.

A. Fish on Hand

Brood Year 2019

Mid-September 2019, LFH had an estimated 109,348 Tucannon River summer steelhead juveniles on hand. 45,040 fish for the conservation program and 64,338 fish for the mitigation program.

B. Tagging, Transfers, and Releases

In September, all Tucannon River endemic steelhead for the conservation portion of the program will be CWT only. The mitigation portion of the program will be 100% ad-clipped with 25,000 also receiving a CWT. In October 2020, the conservation group fish (45,000) will be moved to the TFH where they will be reared until release as yearlings in April or early May from Curl Lake AF. Prior to 2016, releases have been roughly five miles upstream of the TFH, just below the Curl Lake intake structure. Beginning in 2016, WDFW staff began transferring the smolts into Curl Lake after the spring Chinook were released. This release method is expected to continue into the future. The group marked for harvest mitigation will be full term reared at LFH and released at Marengo Bridge in April or early May, depending on size. A total of 15,000 fish will be PIT tagged by Biomark Inc. prior to release (Table 12).

Table 13. Proposed BY19 Tucannon River summer steelhead production.

Site	BY19Goal	Expected	Size	Age	Mark/CWT	PIT	Transfer/Release
		at Release	(fpp)			Tags	Date
Tucannon River at Curl Lake	50,000	45,000	4.5	1+	100% CWT Only	7,500	April 2020
Tucannon River (Marengo Bridge)	100,000	64,000	4.5	1+	25,000 Ad/CWT 39,000 Ad Only	7,500	April 2020

C. Trapping

Brood Year 2020

Trapping of BY20 Tucannon River endemic stock will begin in February (depending on seasonal weather) at the Tucannon FH adult trap (located adjacent to the Rainbow Lake Intake) and is generally completed by mid-May. Tucannon FH staff use EN to calm the fish for handling, transferring only a portion of unmarked natural origin adults and tagged endemic origin adults to LFH based on broodstock needs. All NOR's and endemics not needed for broodstock are released up river of the trap.

Current survival estimates indicate that 35-38 spawned females (depending on age structure) will provide enough eggs to meet the current smolt production goal. WDFW will target 40 to 42 females to be brought to the hatchery for broodstock needs, with up to 14 being NOR. Any females not used will be returned to the river to spawn naturally. Per co-manager agreement, a pre-season estimate based on PIT tag returns on the number natural origin fish expected at the TFH adult trap will be made. Per the Tucannon Steelhead Broodstock Sliding Scale (Appendix G), the appropriate number of natural and hatchery origin fish will be collected for either the conservation or mitigation broodstock.

D. Spawning

Based on fecundity and survival estimates, LFH typically spawns 35-38 females to provide 180,000 green eggs to meet the current conservation and harvest program release goals (Table 13). Spawning occurs in March and April. Matrix spawning is employed due to the relatively small founding population for this program. The intent of this protocol is to spawn two males with each female, increasing genetic diversity and helping ensure successful fertilization of eggs. If not enough males are ripe to achieve this goal; a 1:1 spawning matrix is employed.

E. Rearing

After spawning, fertilized eggs are water hardened in 100 ppm iodophor. They are incubated in down-welling iso-incubation buckets (one fish per bucket) until the eggs eye up. The eyed eggs are shocked, ran through an automated egg sorting machine or handpicked and enumerated, and placed in hatching baskets suspended over shallow troughs. After hatch and swim-up, they are introduced to feed, and transferred to intermediate raceways at around 500 fpp in June. They are transferred again to outside raceways at roughly 200 fpp in July.

Table 14. Proposed BY20 Tucannon River summer steelhead production.

Site	BY20 Goal	Size (fpp)	Age	Mark/CWT	PIT Tags	Transfer/Release Date
Tucannon River (at Curl Lk.)	50,000	4.5	1+	100% CWT Only	7,500	April 2021
Tucannon River (Marengo Bridge)	100,000	4.5	1+	25,000 Ad/CWT 75,000 Ad only	7,500	April 2021

IX. WALLOWA SUMMER STEELHEAD

The Wallowa stock program was initiated to provide a fishery for summer steelhead in the Grande Ronde River (for both Oregon and Washington anglers), and contribute to both tribal, and sport fisheries in the mainstem Columbia and Snake rivers. The overall production of this stock was increased in December 2012, following the elimination of the Lyons Ferry stock steelhead program, and now produces steelhead that are released in the Touchet River from the Dayton AF (100,000), Grande Ronde River from the Cottonwood AF (225,000) and into the Snake River at Lyons Ferry (60,000).

For BY2019, all Wallowa stock steelhead were collected, spawned and early reared by ODFW at Wallowa or Irrigon fish hatcheries in anticipation of the pipeline repairs at LFH.

A. Fish on Hand

Brood Year 2019

Mid-September 2019, LFH had 421,105 Wallowa stock summer steelhead juveniles on hand. After marking, all fish in excess of the needs in Table 2 will be out planted into local lakes.

B. Tagging, Transfers, and Releases

Brood Year 2019

All of these fish were 100% adipose fin clipped into Lake #3 in September, 2019. BY18 was the first group that a single CWT group was used to represent all three release groups, with the CWT fish reared in the lake with the AD only fish (Table 14). A portion of each release group will be PIT tagged just prior to release.

In early February 2020, 225,000 smolts from Lake 3 will be transferred to the Cottonwood AF for final rearing and released into the Grande Ronde River. The fish will be reared/acclimated at Cottonwood AF for approximately 2.5 months and then volitionally released. A total of 6,000 juveniles will be PIT tagged by WDFW for Cottonwood AF prior to release in April; 2,000 of those PIT tags will be used as part of the Comparative Survival Study (CSS) for steelhead production above LGR (Fish Passage Center). After the Touchet spring Chinook are released from the Dayton AF in March, 100,000 will be moved from LFH into Dayton AF. This group will be PIT tagged in the release structure just prior to transfer. They will then remain in the Dayton AF for approximately 2-6 weeks and will be volitionally released during the month of April. The final remaining fish from Lake #3 will be released directly from LFH into the Snake River in mid-April, with a portion being PIT tagged at release in the release structure.

Table 15. Proposed BY19 Wallowa stock summer steelhead production.

Site	BY19 Goal	Expected at	Size (fpp)	Age	Mark/CWT	PIT Tags	Transfer/Release Date
		Release					
Cottonwood AF on the Grande Ronde River	225,000	225,000	4.5	1+	Ad Only Ad/CWT	6,000*	Transfer to Cottonwood AF in Feb from LFH, release in April 2020
Dayton AF on the Touchet River	100,000	100,000	4.5	1+	Ad Only Ad/CWT	4,000	Transfer to Dayton AF in March, released in April 2020.
Snake River (On site at LFH)**	60,000	60,000	4.5	1+	Ad Only Ad/CWT	4,000	On station release in mid-April 2020.

^{*2,000} of these PIT tags are part of the CSS study from the Fish Passage Center

C. Trapping

Brood Year 2020

Trapping of returning Wallowa stock adults occurs on Cottonwood Creek (a small tributary to the Grande Ronde River) March and April. This creek also supplies water to the Cottonwood AF. Because of potential low egg survival and/or IHN virus (both of which have been experienced in recent years), about 110 complete spawned females are needed to provide 475,000 green eggs for the program of 385,000 smolts (Table 15). All unmarked (presumably natural origin) steelhead captured in the Cottonwood Creek adult trap are passed upstream to spawn naturally. All spawned carcasses not considered good quality for food banks will be returned to LFH for burial. If low water flow in the creek does not allow returning adults access to the trap, three alternate strategies may be employed: 1) release juveniles early and begin trapping adults, 2) collection of broodstock at Big Canyon or the Wallowa Hatchery may occur and 3) trap at LFH. Disposition of excess fish (Wallowa Stock HGMP) include 1) killed to collect any CWT fish, 2) offered to local food banks, or 3) killed outright to prevent hatchery swamping of natural origin spawners and hauled to LFH to be buried.

D. Spawning

Spawning generally occurs in late March and early April on a weekly basis. All fish are spawned at the Cottonwood Creek trap site, with the gametes transported to LFH for fertilization, incubation and rearing. A 1:1 male to female mating ratio will continue to be employed whenever possible (see research section below). Adults trapped at LFH or excess adults from ODFW's Wallowa Hatchery or Big Canyon site may be used to provide eggs for this program in the event that enough adults are not collected at Cottonwood Creek.

E. Rearing

After spawning, fertilized eggs are water hardened in 100 ppm iodophor. They are incubated in down-welling iso-incubation buckets (one fish per bucket) until the eggs eye up. The eyed eggs are shocked, ran through an automated egg sorting machine or handpicked and enumerated and placed in hatching baskets suspended over shallow troughs. After hatch and swim-up, they are introduced to feed, and transferred to outside raceways at roughly 500 fpp in June.

Site	BY20	Size	Age	Mark/CWT	PIT	Transfer/Release
	Goal	(fpp)			Tags	Date
Cottonwood AF on the Grande Ronde River	225,000	4.5	1+	Ad Only Ad/CWT	6,000*	Transfer to Cottonwood AF in Feb from LFH, release in April 2021
Dayton AF on the Touchet River	100,000	4.5	1+	Ad Only Ad/CWT	4,000	Transfer to Dayton AF in March, release in April 2021.
Snake River (On site at LFH)	60,000	4.5	1+	Ad Only Ad/CWT	4,000	Direct stream release in mid- April 2021

^{*2,000} of these PIT tags are part of the CSS study from the Fish Passage Center

X. SPOKANE RAINBOW TROUT

Rainbow trout are reared and planted in both southeast Washington and northwest Idaho to meet LSRCP mitigation goals in both states for lost fishing opportunities as a result of construction and operation of the lower Snake River dams. The original LSRCP goal was 93,000 lbs. However, the WDW determined that in stream habitat improvements, equivalent to the cost of producing 7,000 lbs annually of hatchery trout, was a reasonable exchange, and that was implemented in 1983, which reduced the annual production goal to 86,000 lbs for the Snake River Basin. The SE Washington production goal is 79,000 lbs and the NW Idaho production goal is 7,000 lbs. A small, privately funded program (Tri-State Steelheaders, TSS) at the TFH rears rainbow to 1.5 lbs each, providing a unique fishing opportunity in local lakes. This locally funded program replaced the previously state funded program in 2011 which had been in place since the LSRCP took ownership of the Tucannon Hatchery. The agreement at that time was that the state funded program would be allowed to continue at the TFH.

A. Fish on Hand

Brood Year 2018

Mid-September 2019, LFH and TFH had a combined total of approximately 232,868 Spokane stock rainbow trout on hand, this includes diploids and triploids. Fish were shipped to Irrigon

Hatchery in May and shipped back to LFH the week of August 5th. At the time of fish splits and plants in the fall of 2019, staff will determine the exact overage and plant the excess fish into area lakes at the recommendation of fish management. LFC will keep approximately 5% over release goals in the fall to accommodate for mortality and predation.

B. Tagging, Transfers, and Releases

The IDFG fall catchables will be planted in the Moose Creek Reservoir by IDFG staff in late September or early October 2019. All fish for IDFG are triploids from the Spokane stock rainbow trout. Refer to Table 15.

In the spring of 2020, 74,000 catchable diploids (2.5 fpp) and 1,000 jumbos (1.5 lbs. each) will be planted by LFH staff into various lakes in southeast Washington. Spring planting begins in February and is completed in early April.

At the TFH, the goal is to plant 94,000 rainbow trout into various lakes in southeast Washington as catchables (2.5 fpp average.). Planting typically begins in April and is generally completed by the end of June. The jumbo trout from TSS program (usually around 4,000) are planted February through May each year, supplementing catchable plants. No Spokane stock rainbow trout are tagged or fin clipped at LFH or TFH.

Table 17. Proposed BY 2018 Spokane rainbow trout tagging, transfers and plants

Facility	BY18	Expected	Size		Age	Transfer/Release
	Goal	at release	(fpp)	Lbs.		Date
	16,000	16,000	3.0	5,333	1+	Transfer to and planted by IDFG in Sept/Oct 2019
	32,500	32,500	3.0	9,833	1+	Planted in early Oct 2019
Lyons Ferry	74,000	74,000	2.5	29,600	1+	Planted in Feb-Apr 2020
	1,000	1,000	0.67	1,493	1+	Planted in Feb-Apr 2020
	1,650	1,700	1.0	1,650	1+	Transfer to and planted by NPT in Mar-May 2020
Tucannon	95,185	105,348	2.5	38,074	1+	Planted in Mar-June 2020
	4,000*	4,000*	0.7	5,714	1+	Planted in Feb-May 2020

^{*} These fish are funded by TSS.

C. Rearing

Brood Year 2019

Eggs for Washington's legal and jumbo programs, along with Idaho's fall catchable plants come from WDFW's Spokane Hatchery (Spokane stock). WDFW managers completed an Inland Trout Stocking Plan in 2012 for all hatcheries and water bodies in Washington. The management strategy is to plant larger catchables (2.5 fpp) at reduced numbers. Total pounds reared were not affected (Table17).

Approximately 65,000 eyed triploid rainbow eggs for IDFG, NPT and the WDFW Rock Lake fall plant will be transferred from the Spokane Trout Hatchery to LFH in December. After trough rearing, they are transferred to outside standard raceways in March. In January, LFH will receive about 91,500 eyed Spokane diploid rainbow eggs for the balance of its catchable and jumbo program. Early rearing is conducted in either shallow troughs or intermediate raceways before transfer to outside standard raceways in April.

The Tucannon Hatchery will receive about 125,000 eyed rainbow eggs in January. Of these, 94,000 will be destined for planting as catchables (2.5 fpp) and 4,000 are destined for planting as jumbos (1.5 lbs each). After receiving these eggs in January, a small portion (1,750) is transferred from TFH to regional education programs, now privately funded by the TSS club. The catchable program group is started in shallow troughs, intermediate reared in outside round tanks and final reared in the earthen rearing pond. The jumbos start in shallow troughs as well and finish in the round tanks. The entire jumbo program at TFH is privately funded by the TSS organization.

Table 18.	Proposed BY	2019 Spokane	rainbow trout releases.
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Facility	BY19 Goal	Expected at release	Size (fpp)	Lbs.	Age	Transfer/Release Date
	16,000	16,000	3.0	5,333	1+	Transfer to and planted by IDFG in Sept/Oct 2020
	32,500	32,500	3.0	9,833	1+	Planted in early Oct 2020
	74,000	74,000	2.5	29,600	1+	Planted in Feb-Apr 2021
Lyons	1,000	1,000	0.67	1,493	1+	Planted in Feb-Apr 2021
Ferry	1,650	1,700	1.0	1,650	1+	Transfer to and planted by NPT in Mar-May 2021
Tucannon	94,000	105,348	2.5	37,600	1+	Planted in Mar-June 2021
	4,000*	4,000*	0.67	6,119	1+	Planted in Feb-May 2021

^{*}NOTE; Jumbo trout from TSS funding. Total numbers and/or pounds not included in mitigation.

XI. RESEARCH

WDFW (Fish Management or Fish Science staff) are involved in a variety of research, monitoring and evaluation projects throughout SE Washington. Funding of these activities comes from a variety of sources and many are not directly related to the LSRCP Lyons Ferry/Tucannon Annual Operations Plan, but are provided here in general context for the comanagers so they are aware of activities. Some of the below activities are covered under the RM&E Statement of Work submitted to LSRCP under the hatchery evaluation program.

Fall Chinook

- 1) WDFW currently conducts fall Chinook spawning ground surveys in the Tucannon River to document abundance, distribution, and origin of spawners. Coho salmon redds are also estimated during these surveys. (LSRCP)
- 2) WDFW operates a smolt trap on the lower Tucannon River for estimating natural origin salmonid smolt production (spring Chinook, fall Chinook, and summer steelhead). (LSRCP)

3) In 2019, WDFW will again collect fecundity samples from fall Chinook spawned at Lyons Ferry. With PBT being able to distinguish between hatchery and natural origin fall Chinook, a subsample of individual females will be sampled at eye-up so fecundity by origin (hatchery [yearling or subyearling] or natural) can be documented. (LSRCP)

Spring Chinook

- 1) Due to the recent history of high pre-spawn mortality for Tucannon River spring Chinook salmon, it was agreed for 2019 returns that all the fish at the TFH adult trap be kept at LFH for broodstock or adult out planting in late August. Outplant success from the previous years when this was done varied considerably (60-95%). Based on the numbers on hand currently, it's doubtful that outplants will occur in 2019. (LSRCP)
- 2) WDFW currently conducts spring Chinook carcass and spawning ground surveys in the Tucannon River to document pre-spawn mortality, abundance, distribution, and origin of spawners. Surveys are also used to estimate total returns to the river. (LSRCP)
- 3) WDFW operates a smolt trap on the lower Tucannon River for estimating natural origin smolt production (spring Chinook, fall Chinook, and summer steelhead). Annually, up to 5,000 NOR spring Chinook are PIT tagged for juvenile outmigration and adult return monitoring. (LSRCP)
- 4) WDFW conducts spring Chinook redd surveys (as needed) in the Touchet River to document spawning from adult outplants by CTUIR in the North Fork and Wolf Fork of the Touchet River. (BPA, WDFW State Funds)

Summer Steelhead (by basin)

Asotin Creek:

- 1) WDFW operates adult weirs for summer steelhead in the Asotin Creek population. Current trap locations include Asotin Creek, George Creek, and Alpowa Creek. The weirs are used to estimate natural and hatchery origin abundance at all locations, and for collection of biological samples of returning steelhead for population age and genetic structure. (BPA)
- 2) WDFW operates a smolt trap in the mainstem of Asotin Creek (below the Asotin Creek and George Creek weirs) for estimating natural origin smolt production (primarily summer steelhead, but spring/fall Chinook are also captured) from the basin. Annually, up to 3,500 summer steelhead are PIT tagged for juvenile outmigration and adult return monitoring. (BPA)
- 3) WDFW is partially funded by the Asotin Creek Intensively Monitored Watershed (IMW) for juvenile sampling in the upper basin, some hook/line sampling and PIT tagging of summer steelhead, and maintenance/operation of PIT Tag arrays within the Asotin Creek basin that are part of the IMW study. (Pacific Salmon Coastal Fund, WA State Salmon Recovery Fund).

Small Snake River Tributaries:

1) WDFW operates adult weirs (rotating panel selection) of small tributaries located between the mouth of the Tucannon River and Lower Granite Dam. Currently, these tributaries have been designated as part of the Tucannon steelhead population. The weirs are used to estimate natural and hatchery origin abundance at all locations, and for collection of biological samples of returning steelhead for population age and genetic structure. (BPA project #2010-028-00)

Touchet:

- 1) A paired release study was being performed utilizing 10,000 PIT tags to monitor the success of WxW or WxH crosses in the Touchet Endemic steelhead program. Four years of tagging have been completed (BY15 BY18), with adult returns to be finalized in the coming years. Releases of both study groups were occurred from the Dayton Acclimation pond. First year of release was in 2016, final year or release was in 2019, and final adult returns will be completed with the 2021 run year. Results to date indicate a small survival advantage for the WxH group compared to the WxW group (0.50% to 0.45%). (LSRCP, BPA, Walla Walla Monitoring Project)
- 2) WDFW operates a smolt trap on the lower Touchet River for estimating natural origin smolt production (primarily summer steelhead, but spring and fall Chinook have also been captured/documented). Annually, we target 4,000 (or more) summer steelhead to PIT tagged for juvenile outmigration, estimating adult returns, and overshoot monitoring. (WA State Salmon Recovery Fund Fish In/Fish Out Projects, BPA Walla Walla Monitoring Project, LSRCP)
- 3) WDFW operates adult steelhead traps on Coppei and Patit creeks to monitor abundance of natural and hatchery origin spawners, and collection of biological samples for age composition. (Walla Walla Monitoring Project BPA)
- 4) WDFW currently conducts summer steelhead spawning ground surveys in the Touchet River basin (locations above Dayton) to estimate abundance and distribution of spawners. (LSRCP)
- 5) WDFW operates and maintains a series of PIT tag arrays (Harvey Shaw, Bolles, Coppei, Patit, and Dayton), for monitoring adult steelhead (hatchery and wild) returns to the basin. (BPA Walla Walla Monitoring Project, LSRCP)

Tucannon:

- 1) WDFW operates a smolt trap on the lower Tucannon River for estimating natural origin smolt production (spring Chinook, fall Chinook, and steelhead). Annually, we target 2,500 summer steelhead (or more) for PIT tagging for adult return estimation and overshoot monitoring. (LSRCP, BPA PIT Tags for Steelhead only)
- 2) WDFW currently conducts summer steelhead spawning ground surveys in the upper Tucannon River only (and Cummings Creek) to estimate abundance of spawners in correlation with fish passed at the Tucannon adult trap. (BPA)
- 3) WDFW operates and maintains a series of PIT tag arrays (Lower Tucannon, Middle Tucannon, Upper Tucannon, Tucannon FH), for monitoring adult steelhead (hatchery and wild) returns and distribution throughout the basin. Arrays are also used for adult spring Chinook and bull trout monitoring. (BPA, LSRCP)

Grande Ronde:

1) See additional document sent with the 2014/2015 AOP, Wallowa Stock rearing/acclimation study proposed by WDFW and ODFW: A Survival and Straying Comparison of Wallowa Stock Steelhead Reared at WDFW's Lyons Ferry and ODFW's Irrigon Fish Hatcheries. This study is examining survival and stray rate differences from summer steelhead reared at either Irrigon Fish Hatchery or Lyons Ferry Fish Hatchery. Groups from each rearing facility were transferred to either Wallowa Hatchery or Cottonwood Acclimation for release. At Cottonwood, both CWT's and PIT's will be used to evaluate the groups, while at Wallowa Hatchery, just PIT tags will be used for comparisons. To date, only PIT tag returns to Bonneville have been summarized. Overall, survival of LFH reared fish has averaged 2-3 times greater than those reared at Irrigon FH, regardless of release location. To date, two of four years have completed returns, with 1-salt returns for study year three. WDFW will be providing a more complete summary at next year's LSRCP annual meeting with results through the spring of 2020. Differences in stray rates from these groups from either release location have yet to be examined. (LSRCP)

XII. FISH HEALTH

A. Guiding Policies

All fish production at LFH is conducted according to the Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State and Integrated Hatchery Operations Team (IHOT) fish health policy. Specifically, all lots of fish are monitored for fish health, all broodstock are inspected annually, strict hatchery sanitation procedures and fish culture practices (rearing criteria) are followed, and egg and fish transfer and release requirements are met. The attending aquatic veterinarian will respond to all fish disease outbreaks and oversee fish health-related services. Bacterial kidney disease (BKD) management for Chinook stocks, imposed by Idaho and Oregon state agencies' transfer policies, is outlined in Section C.

B. Monitoring

The aquatic veterinarian will regularly visit LFH and TFH to inspect stock, address mortality events, and provide consultation per their discretion and as needed to maintain a veterinary client patient relationship (VCPR). During site visitations, any fish cohorts exhibiting clinical signs, morbidity, and/or abnormal mortality may be examined by the veterinarian; approximately five to ten fish of each species may be sacrificed for necropsy. Updated mortality records, loading

forms, and treatments logs will be provided to the veterinarian upon their request. Copies of fish health reports, veterinary feed directives (VFDs), and prescriptions referencing fish at each respective hatchery will be keep on file on site for at least three years.

At spawning, all broodstock will be tested for viral pathogens. Samples of ovarian fluid from 60 females and kidney/spleen tissue from 60 adults of either sex will be submitted for testing. Samples will be pooled into groups of up to five individual fish. In the event that 60 female fish cannot be obtained, all eligible fish from the population will be sampled (i.e. steelhead stocks for the Tucannon and Touchet). Standard hatchery practices of egg disinfection and use of pathogen-free rearing water during early rearing are considered sufficient fish health measures for the control of viral pathogens, including infectious hematopoietic necrosis virus (IHNV).

To comply with Idaho's fish import regulation, kidney/spleen samples from 60 rainbow trout will be tested for viral pathogens four to six weeks before transfer. Upon completion, results will be communicated to IDFG.

C. Specific Fish Health Management

1. BKD Management – Fall Chinook

Starting with BY16, all females spawned at Lyons Ferry will be 100% tested using ELISA. This is to allow more flexibility in shipping eggs and also for using fry for either yearling or subyearling programs.

WDFW categorizes BKD-ELISA optical densities as follows:

- Below-low = < 0.099,
- Low = 0.099 to 0.198,
- Moderate = 0.199 to 0.45,
- High = > 0.45

Progeny of negative (below low) females will be selected for the yearling fall Chinook program. Eggs from below low and low females will be selected for shipment to the states of Oregon and Idaho. ODFW has agreed to perform the sampling and testing on 150 females at LFH during spawning. WDFW takes the remaining samples, with a portion of the ELISA testing paid for by IPC (350 females/season). Progeny of all low, moderate and high BKD-ELISA females may be utilized in the subyearling fall Chinook program for WDFW releases and Captain John's landing.

2. BKD Management – Spring Chinook

Starting with BY17, all pre-spawning antibiotic injections intended for 1) the control of adult mortalities associated with *Renibacterium salmoninarum* and/or 2) the mitigation of vertical transmission of *R. salmoninarum* to progeny were suspended until an evidence-based risk of disease, confirmed by necropsy findings and appropriate ancillary testing, and which reasonably

threatens the welfare of the broodstock program, was established. Starting with BY18, all female spring Chinook broodstock will receive a pre-spawning injection of tulathromycin (Draxxin®) at a target dose of 2.5 mg/kg in the dorsal sinus upon collection and a second injection of tulathromycin (Draxxin®) at the same dose in the dorsal sinus 30 days prior to spawning. WDFW may go back to Erythromycin with it once again being available for use. Pre-spawning female spring Chinook will be evaluated for BKD-antigen via ELISA assay and indexed according to the resulting value. (Refer to WDFW categorizes of optical densities under section C. 1. "BKD Management of Fall Chinook"). The following culling program is intended to minimize bacterial load within the population and reduce risk of infection: Until the ELISA results are known, eggs from individual females will be incubated separately. Egg reductions required to meet production targets will begin with the highest ELISA range and proceed downward by range until the production target is met. If possible, all eggs from adults with an ELISA value of 0.45 or higher will be culled from the program. If it is determined that all eggs must be kept to meet program requirements, and space is available, groups with ELISA readings above 0.45 may be segregated and reared separately. No segregation between production groups with ELISA readings below 0.45 will occur. Preferably, only the eggs from females with the lowest ELISA values will be kept for the program.

3. Summer Steelhead

At spawning, all broodstock will be tested for viral pathogens. Samples of ovarian fluid from 60 females and kidney/spleen tissue from 60 adults of either sex will be submitted for testing. Samples will be pooled into groups of up to five individual fish. In the event that 60 female fish cannot be obtained, all eligible fish from the population will be sampled (i.e. steelhead stocks for the Tucannon and Touchet). Standard hatchery practices of egg disinfection and use of pathogen-free rearing water during early rearing are considered sufficient fish health measures for the control of viral pathogens, including infectious hematopoietic necrosis virus (IHNV). No culling is planned due to IHNV.

4. Broodstock and Egg Fungus Management

All adult Chinook and steelhead held for broodstock or for adult out planting will be treated with formalin every other day to control external fungus. All eggs will be treated with formalin daily to control fungus. Treatments will be started 24 hours after fertilization. Treatment of Chinook eggs will halt seven days before hatch. Steelhead egg treatments will stop when the eggs are transferred to baskets for hatching.

Rainbow trout eggs are received at the eyed stage and are not treated with formalin.

XIII. COMMUNICATION

The list of people on the following table are either directly involved in the operation of the LFC, or in related programs and facilities.

Name	Agency	Position	Phone	Email
Policy				
Chris Donley	WDFW	Reg. 1 Fish Program Mgr.	509-892-7861	christopher.donley@dfw.wa.gov
Julie Collins	USFWS	LSRCP Project Lead	208-378-5668	julie collins@fws.gov
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Gary James	CTUIR	Fisheries Program Mgr.	541-276-4109	garyjames@ctuir.org
Dave Johnson	NPT	Fisheries Dept. Mgr.	208-621-3736	davej@nezperce.org
Lance Hebdon	IDFG	Anandromous Fish Mgr.	208-334-3791	<u>lhebdon@idfg.idaho.gov</u>
Production				
Ace Trump	WDFW	LFC Manager	509-646-9201	Ace.Trump@dfw.wa.gov
Derek Gloyn	WDFW	LFH Manager	509-646-3454	Derek.Gloyn@dfw.wa.gov_
Doug Maxey	WDFW	TFH Manager	509-843-1430	Douglas.Maxey@dfw.wa.gov
Chris Starr	USFWS	Hatchery Coordination	208-378-5329	chris_starr@fws.gov
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Appendix A: 2019 Requests for Fall Chinook Production Fish/Eggs (2019 Broodyear)

2018-2027 US v Oregon	Who	Release site	Age	# for release	transfer	Survival to release or transfer (revised 6/22/16)	Expanded for loss prior release (1/F)		SRL Calcs	Total estim eggtake which will cover needs through this priority
1	WDFW	onstation	yearlings	450,000		94.2%	1.06123	477,555	94.2% mean survival BY11-15	477,555
		total yearlings	450,000							
2	NPT	CJ	subs	450,000	453,492	94.9%	1.05383	474,223	99.23% survival transfer to rel BY11-15	951,778
3	NPT	вс	subs	450,000	453,492	94.9%	1.05383	474,223	3.1% green to eye, est 2% eye-transf, 1% transf-rel	1,426,002
4	WDFW	onstation	subs	500,000		94.4%	1.05988	529,942	94.4% mean survival BY11-15	1,955,943
5	NPT	PIT	subs	400,000	403,104	94.9%	1.05383	421,532	99.95% surv transf to rel BY13-15	2,377,475
6	NPT	CJ2	subs	200,000	200,100	94.9%	1.05383	210,766		2,588,241
7	NPT	BC2	subs	200,000	200,100	94.9%	1.05383	210,766		2,799,007
8	NPT	PIT2	subs	200,000	200,100	94.9%	1.05383	210,766		3,009,773
11	WDFW	onstation	subs	200,000		94.4%	1.05988	211,977		4,574,175
		total subyearlings	2,600,000						2,618,231	<u> </u>
9	WDFW/Irrigon	Salmon R	eyed eggs	1,000,000	1,100,000	96.9%	1.03199	1,135,191	3.1% green to eye loss at LFH (BY11-BY15)	4,144,964
10	WDFW/Irrigon	GRR-direct rel	eyed eggs	200,000	210,500	96.9%	1.03199	217,234	requested numbers by IPC and Irrigon	4,362,198
		total eyed eggs	1,200,000						1,352,425	i
			4,250,000	released				4,574,175	green eggs to meet needs through priority 11	
		number of Snake River orig	gin females needed to s	pawn				1236	(Estimated using 3700 eggs/F), BY16-18 LGR, LFH (3,820 eggs/F)	
		Female trapping goal to	meet requests throu	gh priority 11	l:			1323	(approx 5% strays, 2% mortality, non-viable)	

Appendix B: 2019 Fall Chinook Trapping/Sampling Protocols at LGR

July 26, 2019

Protocols:

- 1) These protocols presume a 24 hour/day, 7 days per week trapping at 70%. Fish trapped during a 24 hour 7 day a week trapping period will not be operculum punched. If the systematic sampling rate is changed, all fish hauled to hatcheries must receive an operculum punch on the right side (ROP) and if trapping changes to only 4 hours per day (100%trap rate), all fish hauled to the hatcheries must receive an operculum punch on the left side (LOP).
- 2) Males and females will not be inoculated.
- 3) All fish \geq 67 cm will be hauled to LFH and NPTH. LFH will haul 70% and the NPT will haul 30%.
- 4) Wire tagged males <70 cm hauled to LFH.
- 5) Wire tagged females <70 will be hauled to LFH and NPTH under the normal 70/30 split. Based on water quality conditions, fish hauled for NPT brood may be held at LFH.
- 6) Unmarked/untagged females < 70 will be hauled to LFH.
- 7) Jacks suspected of being summers will need to be subsampled for wires.
- 8) Only scale sample fish released from the trap. Do not scale sample hauled fish.
- 9) DNA sample all fish trapped regardless if hauled to the hatchery or released.

Wire tagged fish:

Fork Length	Action
≥ 70cm	Haul all wires (DNA sample all)
	Haul 1 out of 4 wires (put F in with "LARGES" for LFH and NPT and
<70 cm	M go into tank for LFH), DNA sample all
	Release 3 out of 4 wires (DNA sample all)
Untagged fish :	
Fork Length	Action
$\geq 70 \text{ cm}$	Haul all fish (DNA sample all).
	Haul 1 out of 4 F to LFH (DNA sample all).
	Release 3 out of 4 F (collect scales and DNA).
<70 cm	Release all M (collect scales and DNA).

Appendix C: 2019 Trapping, Mating, and Sampling Protocols at LFH

LFH may start up the volunteer trap if a shortfall of females or males being collected at LGR happens. If volunteer trap is started, staff will try to high grade for larger adults.

Sorting protocol

Count and sex all fish: 1) Males and females.

Count LGR trapped females returned to the pond during the spawn day.

Sampling protocol

LFH staff processing DIPS: Document Fork length, sex, presence/absence of CWT, and PIT tag number. Take scales and take the snout of the fish if CWT is detected.

SRL staff processing during spawning days:

Processing table: Fin clips for DNA: take sample on every fish so data can be used for run reconstruction purposes, as well as profiling broodstock.

Scales: taken on all fish

Female broodstock total body weights

1st week of spawning: weigh first 50 females that have a CWT and the first 50 females that are unmarked/untagged (appear wild) and note fish ID number

 2^{nd} week- 4^{th} week: weigh first 25 females that have a CWT and 25 females that are unmarked/untagged each spawn day

Carcasses for nutrient enhancement: After otoliths are taken from the carcasses, a tote of fish will be filled and dumped into a bin next to the loading dock. These fish will be frozen separately and taken to the Tucannon River for nutrient enhancement after ELISA testing.

Mating protocol at LFH

Our goals are to maximize the use of potentially natural origin fish and larger/older aged fish and to exclude jills and strays from broodstock.

All wire tagged males must wait until their CWTs are decoded before they are used in a mating.

Stray males will be culled based on CWTs. If broodstock limited, up to 56 stray females may be spawned and retained, presuming 1,112 matings are needed to make production. Any male used on a stray female must also be used on another female that will be retained for production (inbasin hatchery origin, or untagged unknown origin).

Wire tagged males verified as adults can be used on multiple females, with a goal of one male to two females.

Untagged Males \geq 75 cm can be used on multiple females.

Untagged Males 70-74 cm will only be used in 1 x 1 crosses unless there is a shortage of males.

Males <70 cm will not be used in matings unless they are verified as adults. This size criteria may be adjusted in season.

Fecundity monitoring and Jills

All females will be spawned when ripe and the gametes will be held in incubators until we can determine if we have enough adult females to offset the culling, and to monitor fecundity. If we have enough adult females to make production goals, after eye up and fecundity estimation, jills will be culled. Jills verified by CWTs will be spawned with males of a larger fork length. Any male used on a jill must also be used on a larger or older aged fish that will be retained for production. This will be done to ensure if the jill is culled or a fry plant is made, the gametes from the male will still contribute elsewhere in production.

Appendix D: FCAP Facilities

1.1 Pittsburg Landing

The acclimation facility at Pittsburg Landing consists of: 16 -20ft aluminum circular tanks; 2 aluminum distribution boxes; 4 river intake screens; ring lock flexible hose: 4" = 1,260 ft, 6" = 1,780 ft, 8" = 3,110 ft; camlock flexible hose: 6" = 2,080 ft; 1 - 500 gallon diesel storage tank; 1 - 20ft storage container; 2 - 30ft camp trailers; 1 - 1996 Chevy S-10 pickup; two alarm systems; 16 emergency oxygen systems - hoses, micro diffusers and regulators (1 per tank); a trailer mounted 4,000 watt generator light plant; one utility storage trailer; 16 camouflage nets; 2 trailer mounted hydrocyclones; miscellaneous bolts, seals, camlock fittings, etc. Equipment used at Pittsburg Landing and the other two facilities was purchased by USCOE, Walla Walla under the FY95 Congressional Add-on (Senate Report, 103-672, p7).

Water is pumped directly from the Snake River to the acclimation tanks by four, 4-inch electric pumps powered by diesel generators. Generators are rented from a contractor because leasing appeared to offer the least cost over a ten-year life cycle. Each pump has a portable water intake screen that is placed into the river each year and connected to the pump by 120 ft of 6-inch plastic hose. The pumps provide 500 gpm of water and operate 24 hours each day throughout the 6-week acclimation period except for oil checks and servicing. A 1,000 gallon tank, placed within a spill containment barrier, supplies fuel for the pumps. The water is pumped to one of two12 ft. high water distribution boxes, containing degassing towers to remove nitrogen gas, before flowing through a series of downsizing pipes to the rearing units.

The rearing units consist of 16 circular aluminum tanks, 20 ft in diameter and 4 feet deep. The tanks are transported from the storage area by a 20 ft flatbed lift-truck and placed on leveled 6-inch by 6-inch wood timbers. The tanks, made in two pieces and bolted together, drain water from the center of the tank through an 8-inch pipe placed in a plywood manhole running under the tank. The tank is fitted with vertical 12-inch circular perforated aluminum screen and the water depth controlled by a 6-inch center PVC standpipe.



The rearing water enters the tank through a 4-inch pipe located on the edge of the tank and is directed in a manner to facilitate a circular motion to aid the movement of fish waste and mortality to the center screen. Water flow is controlled by a 4-inch gate valve located on the incoming line and maintains flows at 100 gpm. The water discharge line is connected from the tank to the river by an 8-inch flexible plastic pipe, which is also used to release the fish.

A 24-volt alarm system constantly monitors water levels in each rearing tank and each of the two water distribution towers. An enunciator panel that provides a visual and audio alarm when a low water level is detected monitors the alarm system. The alarm control box and enunciator panel is located near the staff-housing trailer.

Assembly of the acclimation site begins in February each year with the transport of equipment and material from an offsite storage area. In 2006, the U.S. Forest Service (USFS) agreed to a trial operation of allowing the NPT to leave half of the assembled fish rearing tanks in place and remove the other half and related equipment at a storage site near the fish acclimation site. This agreement should greatly reduce equipment fatigue and reduced assembly and disassembly time by half.

1.2 Big Canyon

The Big Canyon facility uses identical or similar equipment to that of Pittsburg Landing. The rearing tank assembly has been changed over the years to include a single row of tanks that sit flat on the gravel surface. The center drain line is located in a trench dug under the tank, thus eliminating the need for 12-inch deep gravel pad that was previously used. This method can only be used where the proper elevation is available to facilitate water discharge to the river.

The USCOE agreed to furnish electric pumps to replace the diesel units that were rented each year. Electric pumps were installed and tested before the 2002 acclimation season. The electric pumps provide the same performance as the diesel pumps while reducing rental and maintenance costs, allowing onsite staff reduction and eliminates the risk of a major fuel spill.

FCAP Project Leader received verbal agreement from the Nez Perce Tribe that allows the fish rearing tanks and water distribution tower to remain assembled at the site the entire year. This eliminates the need for an assembly and disassembly contract and reduces equipment fatigue hence provide dollar savings to the program.

1.3 Capt. John Rapids

The Capt. John Rapids Fall Chinook Acclimation Facility is a single 150 \(\simex\) X50 \(\simex\) in-ground, lined pond that is supplied with Snake River water by two independent 1,250 gpm submersible electric pumps. Other facility equipment and capital construction consists of: 2 river intake screens; one camp trailer; one standby propane generator; one water well (domestic water); septic system; commercial electric service; alarm system; telephone service. The pumps and intake screens were designed to be placed into the river and then removed following fish acclimation each year but were replaced in 2001 with permanent intake screens located in the main Snake River channel. The pump intake screens are provided with an air back flush system to remove debris and an alarm system is available to monitor flows.

The pumps deposited large amounts of sand in the acclimation pond, which was removed by hand tools between each group of fish. The deposited sand created extremely poor environmental conditions for the fish during release

Negotiations with the USCOE resulted in the installation of two sand separators, two larger sized water pumps, and upgrade of the electrical and pump control panels and changes in the pond water alarm system. Installation of the new equipment began in the fall of 2007 and testing indicated that the sand separators removed most of the sand load that had been deposited in previous years.

2. Operations

2.1 Fish transport

Approximately 500,000 sub-yearlings will be transferred to the Big Canyon and CJR facilities and 400,000 will be transferred to Pittsburg Landing. CJR sub-yearlings will be transported by WDFW, while Pittsburg Landing and Big Canyon transports will be shared by NPT and WDFW. Big Canyon and Captain John Rapids sub-yearlings will be transferred in late April while Pittsburg landing sub-yearlings will be transferred around the third week in April. Lyons Ferry Hatchery personnel provide schedules and facilitate loading and enumeration of the fish. Fish transport permits will be requested from IDFG.

The final release of yearlings was in 2019 as the program has shifted to a sub-yearling only release with two groups being released at each facility. The second release group of approximately 200,000 sub-yearlings will be transferred to all three facilities approximately 4-5 days after the release of the first group.

2.2 Rearing

During acclimation, staff perform daily scheduled fish culture duties that includes: checking and recording oxygen levels in the rearing units three times each day, feeding the rearing units three times each day and picking fish mortality twice each day. Staff also observes fish behavior for abnormalities and assist in fish health checks and the fish-marking program. The fish are fed Bio-Clark's fry and Bio-Pro, manufactured by Bio-Oregon of Longview Washington. Fish culture methods are the same as per Integrated Hatchery Operations Team (IHOT) guidelines and consistent with WDFW fish culture techniques at Lyons Ferry Hatchery. Environmental precautions are necessary to handle diesel and oil for the portable water pumps.

Fish health services are provided by contract with the USFWS, Dworshak Fish Health Center (DFHC). The contract provides diagnostic and pathogen survey services for all fall Chinook juveniles and smolts transported to the fish acclimation facilities. The services include a fish health check before transfer, and a pre-release exam. Other health checks are performed as requested. Fish health protocols are as per AFS Blue Book, IHOT and Nez Perce Tribe fish health protocols.

2.3 Marking

Sub-yearling fish will be marked with coded wire tags (CWT), adipose fin clipped and pit tagged prior to transfer to the FCAP facilities.

2.4 Release

Sub-yearling fish are acclimated approximately three weeks before release in early to mid May at 50fpp. The second group of sub-yearlings will be released in late May to early June, also at 50 fpp. Anticipated sub-yearling transfer and release dates for 2020:

- Pittsburg Landing Transfer April 20 Release May 7
- Pittsburg Landing(2) Transfer May 11 Release May 27
- Captain John Rapids Transfer April 27 Release May 13
- Captain John Rapids(2) Transfer May 18 Release June 4
- Big Canyon Transfer April 27 Release May 14
- Big Canyon(2) Transfer May 18 Release June 5

Emergency low water, water temperatures or facility equipment failure may necessitate an early release of fish from the facilities. The facility operator is authorized to determine when to release the fish if emergency circumstances warrant. Co-management agencies will be contacted within 24 hours with notification of an early release.

2.3 Communication

Verbal communications between FCAP personnel and co-managers is done on an as needed basis to facilitate planning, transportation and acclimation. Co-managers will be involved in any planned deviation to the fish acclimation schedule.

Fish release numbers will be reported and a FCAP fish acclimation summary will be completed by Nez Perce Tribe Research division. FCAP fish acclimation summary and other pertinent information will be presented to co-managers at the Snake River Fall Chinook Technical Group meeting.

FCAP personnel will complete and submit a project annual report to LSRCP in August each year.

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Appendix E: 2019 Releases - Fall Chinook Pit Tag Allocation (USvOR agreement)

Summary of PIT tag allocation for release year 2019 Snake River fall Chinook salmon hatchery production.

Summa	ly of i i i tag a	ilocation i	OI IC	lease year 2019 Snake	Kivei lali Cillic	ok samon natene	I production.		
						Release numbers avai	lable for PIT tagging	Tagging Load /	
Priority			Prod	luction Program			Subyearlings		Tagging Lead / Uploading
					PIT Tag #'s	PIT Tag #'s	Yearlings		
	Rearing Facility	Number	Age	Release Location(s)	Monitor Mode	Bypass if Collected	Subyearling Sample Size		
						BIC	Representative Allocation		
1	Lyons Ferry	450,000	1+	On station	$10,000^1$	0		WDFW/WDF	W(monitor mode for SARs)
2	Lyons Ferry	450,000	0+	Capt. John Rapids 1	18,200 ²	$7,800^2$			NPT/NPT
3	Lyons Ferry	450,000	0+	Big Canyon 1	$7,700^2$	$3,300^2$			NPT/NPT
4	Lyons Ferry	500,000	0+	On Station	20,0001			WDFW/WDF	W(monitor mode for SARs)
5	Lyons Ferry	400,000	0+	Pittsburg Landing 1	18,200 ²	$7,800^2$			NPT/NPT
6	Lyons Ferry	200,000	0+	Captain John Rapids 2		$4,500^3$			NPT/NPT
7	Lyons Ferry	200,000	0+	Big Canyon 2		4,500 ³			NPT/NPT
8	Lyons Ferry	200,000	0+	Pittsburg Landing 2		$4,500^3$			NPT/NPT
9	Irrigon	1,000,000	0+	Salmon River		4,5001			IPC/IPC
10	Irrigon	200,000	0+	Grande Ronde		4,5001			WDFW/WDFW
11	Lyons Ferry	200,000	0+	On Station					
NPTH 1	NPTH	500,000	0+	NPTH		$4,500^3$			NPT/NPT
NPTH 2	NPTH	200,000	0+	Lukes Gulch		$4,500^3$			NPT/NPT
NPTH 2	NPTH	200,000	0+	Ceder Flats		$4,500^3$			NPT/NPT
NPTH 3	NPTH	500,000	0+	North Lapwai Valley		$4,500^3$			NPT/NPT
TOTAL	Yearlings			450,000			0	PIT Yearlings	PIT Subyearlings
	Subyearlings			5,200,000				10,000	123,500

Total PIT tags:

LSRCP tags 39,000
 FPCC 63,000
 BPA tags 21,500

HOR = Hatche	ery origin re	eturn					modified	based on T	ribal comm	ents on Jan	ı 18, 2013	-corrected for	mulas for 65%	of run at trap			
NOR = Natura	ıl origin retu	ırn			MAT = 750,	so 555 N	OR at trap	provides ~7	50 NOR to	river							
	icted HOR =		at trap		dict. HOR=	312					Dispostio				PNI Prior	to harvest	
	icted NOR =		at trap	Total pre	dict. NOR=	78		NOR	NOR	HOR	HOR	Program	Tribal & Nonti			pNOB=	
Tot. Est Retur		254	at trap					Brood	SpEsc	Brood	SpEsc	Size	Harvest	Transfer	PNI	pNOS=	
Total River Ret		391	w/ 35% below	trap		391		50	23	106	175	225,000	0	0	0.27	pHOS=	0.88
	ood Target =									w/o harv	est or trans						
•	ed Adult rur	ı size at	the TFH trap i									Total Escap	Total NOS	pNOS	Total HOS	pHOS	
Predicted			PNOB	At trap	At trap	At Trap	At trap	NOR	HOR				Escapement	(NOS	Escapement		
NOR	NOR	HOR	(NOR	NOR	HOR	Total	NOR	total river	total river	Escapem	Size	prespawn	after 15%	escapement		escapemen	ıt
at Trap	Broodst.	Broodst.		SpEsc	SpEsc	Esc	Esc%	escapem.		NOR +HOR		loss	presp loss	%)	presp loss	%)	
50	50	106	32%	0	97	97	0.0%	27	206	233	389			11.5%	175	88.5%	
100	50	106	32%	50	97	147	34.0%	104	206	310	466		88	33.5%	175	66.5%	
150	75	81	48%	75	122	197	38.1%	156	231	387	543	329	132	40.2%	197	59.8%	
200	85	71	54%	115	132	247	46.6%	223	241	464	620	394	189	48.0%	205	52.0%	
250	85	71	54%	165	132	297	55.6%	300	241	541	697	460	255	55.4%	205	44.6%	
300	100	56	64%	200	147	347	57.6%	362	256	618	774	525	307	58.5%	218	41.5%	
350	110	46	71%	240	157	397	60.5%	428	266	695	851	591	364	61.7%	226	38.3%	
400	125	31	80%	275	172	447	61.5%	490	281	772	928	656	417	63.5%	239	36.5%	
450	140	16	90%	310	187	497	62.4%	552	296	849	1005	721	469	65.1%	252	34.9%	
500	156	0	100%	344	203	547	62.9%	613	312	926	1082	787	521	66.3%	265	33.7%	
550	156	0	100%	394	203	597	66.0%	690	312	1002	1158	852	587	68.8%	265	31.2%	
600	156	0	100%	444	203	647	68.6%	767	312	1079	1235	917	652	71.1%	265	28.9%	
650	156	0	100%	494	203	697	70.9%	844	312	1156	1312	983	717	73.0%	265	27.0%	
700	156	0	100%	544	203	747	72.8%	921	312	1233	1389	1048	783	74.7%	265	25.3%	
750	156	0	100%	594	203	797	74.5%	998	312	1310	1466	1114	848	76.2%	265	23.8%	
800	156	0	100%	644	203	847	76.0%	1075	312	1387	1543	1179	914	77.5%	265	22.5%	
850	156	0	100%	694	203	897	77.4%	1152	312	1464	1620	1244	979	78.7%	265	21.3%	
900	156	0	100%	744	203	947	78.6%	1229	312	1541	1697	1310	1044	79.7%	265	20.3%	
950	156	0	100%	794	203	997	79.6%	1306	312	1618	1774	1375	1110	80.7%	265	19.3%	
1000	156	0	100%	844	203	1047	80.6%	1382	312	1695	1851	1441	1175	81.6%	265	18.4%	
1100	156	0	100%	944	203	1147	82.3%	1536	312	1849	2005	1571	1306	83.1%	265	16.9%	
1200	156	0	100%	1044	203	1247	83.7%	1690	312	2002	2158	1702	1437	84.4%	265	15.6%	
1300	156	0	100%	1144	203	1347	84.9%	1844	312	2156	2312		1567	85.5%	265	14.5%	
1400	156	0	100%	1244	203	1447	86.0%	1998	312	2310	2466	1964	1698	86.5%	265	13.5%	
1500	156	0	100%	1344	203	1547	86.9%	2152	312	2464	2620	2094	1829	87.3%	265	12.7%	

Model Calculations and Assumptions						
Cell C5 - Predicted HOR at Tucannon FH trap: This is an entered number based on presea	son projection					
Cell C6 - Predicted NOR at Tucannon FH trap: This is an entered number based on presea	son projection					
Cell C7 - Total Estimated Run at the trap: Sum of HOR and NOR preseason projections						
Cell C8 - Total run at trap divided by 0.65 to estimate total return to Tucannon River, includi	ng downstrear	n of trap (3	5%).			
Cell C9 - Brood Number: This is a constant number of 170 based on a 225,000 production	level at HOR le	evels >500	- would be a	djusted dov	nward at lowe	r HOR levels
Column A - Predicted NOR at the trap: Lookup value column based on cell C6 preseason p	rojection at tra	p.				
Column B - NOR Broodstock Requirement: generally 50% of brood need up to 350 NOR pr	edicted return	to trap, exc	ept at NOR	< 100		
Column C - HOR Brood Requirement: Total brood need - NOR brood						
Column D - NOR Brood Percent: % NOR in broodstock						
Column E - NOR at trap minus NOR broodstock taken						
Column F - HOR Spawning Escapement above the trap after broodstock taken						
Column G - Total NOR and HOR Spawning Escapement above the trap: after broodstock						
Column H - % NOR Escapement Percent above trap						
Column I - NOR total spawning escapement (trap passage plus 35% below the trap)						
Column J - HOR total spawning escapement (trap passage plus 35% below trap)						
Column K - Sum of NOR and HOR total spawning escapement (trap passage plus 35% belo	w trap)					
Column L - Total return to the Tucannon River, including all broodstock taken						
Column M - Total escapement in the Tucannon River after broodstock collection, minus 15%	% prespawn m	ortality				
Column N - NOR escapement after broodstock collection and 15% prespawning loss in rive	r					
Column O - HOR escapement after broodstock collection and 15% prespawning loss in rive	r					

Appendix G: Tucannon River Summer Steelhead Broodstock and Weir Management Sliding Scale.

Estimated NOR Return to Weir (based on PIT Tag Estimates)		Total NOR & HOR needed for Conservation		Conservation Brood		Total Broodstock Needed for	Mitigati	Mitigation Brood		# of HOR's Used for Total Broodstock	NOR's Released Above Weir	
Estill	nates)	Broodstock	Program	NOR	HOR	HOR Program NOR HOR		HOR	Broodstock needs	needs	Min	Max
	<50	78	26	16	10	52	0	52	16	62	10	34
50	200	78	26	18	8	52	0	52	18	60	37	187
201	400	78	26	21	5	52	0	52	21	57	185	384
401	600	78	26	26	0	52	5	47	31	47	378	577
601	800	78	26	26	0	52	10	42	36	42	574	773
801	1000	78	26	26	0	52	15	37	41	37	770	969

Table Continued.....

Estimated NOR Return to Weir (based on PIT Tag Estimates)		Actual HOR Returns		Total Fish Released Above Weir		%NOR Used in Broodstock from Total Return to Weir		% NOR in Conservation Program	% NOR in Total Program	pHOS (effective) above the Weir		PNI above the Weir	
Estin	nates)	Low	High	Min	Max	Min	Max	PNOB	PNOB	High	Low	Low	High
	<50	68	313	78	347	NA	32%	62%	21%	0.96	0.58	0.39	0.51
50	200	70	315	107	502	36%	9%	69%	23%	0.86	0.21	0.45	0.77
201	400	73	318	258	702	10%	5%	81%	27%	0.55	0.12	0.6	0.87
401	600	83	328	461	905	8%	5%	100%	40%	0.38	0.09	0.73	0.92
601	800	88	333	662	1106	6%	5%	100%	46%	0.29	0.07	0.78	0.93
801	1000	93	338	863	1307	5%	4%	100%	53%	0.23	0.06	0.81	0.94

¹⁾ F1 hatchery origin fish - from the conservation program - will be used for broodstock needs in both conservation and harvest programs.

²⁾ No AD-clipped fish will be used for broodstock, though some will be passed upstream to meet the maximum hatchery fish upstream of the weir (375-broodstock needs)

³⁾ Goal is to have about 300-350 total hatchery origin fish (of either group - conservation preferred over mitigation) above the weir - to ensure future broodstock needs

Appendix H. Numbers of PIT Tags and Coded-Wire Tags implanted into spring Chinook, fall Chinook, or summer steelhead at Lyons Ferry Hatchery Complex, and funding source of those tags.

Species	Stock	Age	Release Location	Program Release Goal	PIT Tag (LSRCP)	PIT Tags (BPA)	PIT Tags (Other)	CWT (LSRCP)	CWT (BPA)	PBT Baseline Evaluation
SPCHK	Tucannon	1+	Tucannon River	225,000	7,500	7,500		225,000		BPA
SPCHK	Carson	1+	Touchet River	250,000	15,000			80,000		BPA
FACHK	Snake River	1+	Snake River, Lyons Ferry FH	450,000	10,000			450,000		BPA
FACHK	Snake River	0	Snake River, Lyons Ferry FH	700,000	20,000			200,000		BPA
FACHK	Snake River	0	Snake River, Pittsburgh Landing 1	400,000		4,500		200,000		BPA
FACHK	Snake River	0	Snake River, Pittsburgh Landing 2	200,000		4,500		200,000		BPA
FACHK	Snake River	0	Snake River, Captain Johns Landing1	450,000		4,500		200,000		BPA
FACHK	Snake River	0	Snake River, Captain Johns Landing 2	200,000		4,500		200,000		BPA
FACHK	Snake River	0	Clearwater River, Big Canyon 1	450,000		4,500		200,000		BPA
STL	Wallowa	1+	Touchet R. @ Dayton Acclimation	100,000	4,000			40,000		BPA
STL	Wallowa	1+	Snake River @ Lyons Ferry Hatchery	60,000	4,000			for all three		BPA
STL	Wallowa	1+	Grande Ronde R. @ Cottonwood Pond	225,000	4,000		2,000	groups combined		BPA
STL	Touchet Endemic	1+	Touchet R., Dayton Acclimation	50,000	5,000			50,000		BPA
STL	Tucannon-Mitigation	1+	Tucannon River (lower)	100,000		7,500			25,000	BPA
STL	Tucannon- Conservation	1+	Tucannon River, Curl Lake	50,000		7,500			50,000	BPA
SPCHK	Tucannon	1+	Tucannon River at Smolt Trap	NA	5,000			NA		NA
STL	Tucannon	1+	Tucannon River at Smolt Trap	NA		2,500		NA		NA